# Extracting Biological Information from Gene Lists

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## Programme

- The theory and practice of gene set enrichment
- Gene set enrichment practical
- Presenting results
- Dealing with artefacts and biases
- Motif analysis
- Motif analysis practical

## Standard Gene List Output

Rank	Well	Gene Name	P-value	<b>GPR Fold Change</b>	<b>GPR Fold Change Graph</b>	Control 1	Control 2	Control 3
1	E10	Tnfrsf18	0.006689	-7.708526		26.291138	25.415058	25.808804
2	E08	Ly9	0.009059	-7.238955		25.672344	24.660522	24.845451
3	E01	<u>Tollip</u>	0.081636	-14.769324		27.33491	31.586285	27.811256
4	H10	Stat3	0.092269	-2.377623		25.84287	26.284285	26.874344
5	F03	Nt5e	0.097510	-1.511391		25.420982	26.977015	25.08718
6	C01	Tnfrsf1b	0.099746	-4.026225		40.0	37.44099	36.49696
7	A09	Ccnd3	0.100523	3.755167	<u> </u>	30.837646	30.475822	30.468536
8	D11	Nfatc2	0.124354	5.534758		28.610485	29.669998	30.464863
9	D05	<u>Il2ra</u>	0.132781	-1.549923		37.23	35.44099	35.49696
10	H04	Sema4a	0.133853	-5.447223		36.48277	36.928036	32.373432
11	D01	Tnfsf4	0.144796	6.022623		21.888157	20.845629	22.976254
12	B10	Nfat5	0.145780	8.067699		30.449022	30.795446	30.850525
13	D07	Cd3e	0.166966	5.300400		28.893595	28.981432	30.581322
14	H05	Nrp1	0.171774	3.802116	<u> </u>	30.856043	30.58041	30.099209
15	G05	Cd53	0.180716	-2.249306		33.33491	33.586285	33.811256
16	D09	Cd28	0.188418	-4.313547		24.510563	23.23	20.464325
17	D02	Pou2af1	0.199099	2.734895	•	26.449022	22.795446	23.850525
18	F09	Gadd45b	0.209415	-1.859485		25.837646	25.475822	24.468536
19	D04	S100a6	0.221836	-1.869103		22.482086	24.83037	23.917696
20	B07	Stat6	0.233153	-1.493636	1	33.44925	32.16483	32.71563

## Descriptions aren't always informative

Gene	Description
Gpr55	G protein-coupled receptor 55 [Source:MGI Symbol;Acc:MGI:2685064]
Ncl	nucleolin [Source:MGI Symbol;Acc:MGI:97286]
Aspm	asp (abnormal spindle)-like, microcephaly associated (Drosophila) [Source:MGI Symbol;Acc:MGI:1334448]
Tnfsf4	tumor necrosis factor (ligand) superfamily, member 4 [Source:MGI Symbol;Acc:MGI:104511]
Ephx1	epoxide hydrolase 1, microsomal [Source:MGI Symbol;Acc:MGI:95405]
Setx	senataxin [Source:MGI Symbol;Acc:MGI:2443480]
Angptl2	angiopoietin-like 2 [Source:MGI Symbol;Acc:MGI:1347002]
Ggta1	glycoprotein galactosyltransferase alpha 1, 3 [Source:MGI Symbol;Acc:MGI:95704]
Dab2ip	disabled homolog 2 (Drosophila) interacting protein [Source:MGI Symbol;Acc:MGI:1916851]
Neb	nebulin [Source:MGI Symbol;Acc:MGI:97292]
Ermn	ermin, ERM-like protein [Source:MGI Symbol;Acc:MGI:1925017]
Ckap5	cytoskeleton associated protein 5 [Source:MGI Symbol;Acc:MGI:1923036]
Prr5l	proline rich 5 like [Source:MGI Symbol;Acc:MGI:1919696]
Arhgap11a	Rho GTPase activating protein 11A [Source:MGI Symbol;Acc:MGI:2444300]
Bub1b	budding uninhibited by benzimidazoles 1 homolog, beta (S. cerevisiae) [Source:MGI Symbol;Acc:MGI:1333889]
Prnp	prion protein [Source:MGI Symbol;Acc:MGI:97769]
Fam102b	family with sequence similarity 102, member B [Source:MGI Symbol;Acc:MGI:3036259]

## Gene summary sites are useful for single genes

<u>TNFSF4</u> - tumor necrosis factor (ligand) superfamily...

Homo sapiens

Synonyms: CD134L, CD252, GP34,

Glycoprotein Gp34, OX-40L, ...

Biagi, E. et al., Godfrey, W.R. et al., Wang, X. et al., Takasawa, N. et al., Ito, T. et al., et al.

Welcome! If you are familiar with the subject of this article, you can contribute to this open access knowledge base by deleting incorrect information, restructuring or completely rewriting any text. Read more.

#### Disease relevance of TNFSF4

- In two independent human populations, the less common allele of SNP rs3850641 in TNFSF4 was significantly more frequent (P <or= 0.05) in individuals with myocardial infarction than in controls [1].
- However, cytotoxic T lymphocyte (CTL) clones specific for Epstein-Barr virus
   (EBV)-transformed autologous lymphoblastic cell lines (LCLs) induced both
   OX40 and OX40L expression after antigen or T cell receptor (TCR)
   stimulation [2].
- We have cloned and sequenced a cDNA encoding gp34, a novel glycoprotein expressed in cells bearing human <u>T-cell</u> leukemia virus type I (<u>HTLV-I</u>) [3].
- On the other hand, gp34 was not expressed on these cells, although its expression is also known to be associated with <a href="https://https://https://html.ncbi.nlm.ncbi.
- Regulation of <u>T cell</u> activation <u>in vitro</u> and in vivo by targeting the OX40-OX40 ligand interaction: amelioration of ongoing <u>inflammatory bowel disease</u> with an OX40-IgG fusion protein, but not with an OX40 ligand-IgG fusion protein [5].

#### High impact information on TNFSF4

- We therefore conclude that Tnfsf4 underlies <u>Ath1</u> in mice and that polymorphisms in its human homolog <u>TNFSF4</u> increase the risk of <u>myocardial</u> <u>infarction</u> in humans [1].
- The <u>quantitative trait locus</u> region encompasses 11 known genes, including Tnfsf4 (also called <u>Ox40</u>l or Cd134l), which encodes OX40 ligand [1].
- When activated in the presence of leukemic CLL <u>B cells</u>, <u>T cells</u> rapidly upregulate CD30 through an <u>OX40 ligand</u> and <u>interleukin 4</u> (IL-4)-dependent mechanism [6].
- Here we report that <u>TSLP induced</u> human DCs to express <u>OX40 ligand</u> (OX40L) but not IL-12 [7].
- <u>TSLP-induced OX40L</u> on DCs was required for <u>triggering</u> naive CD4(+) <u>T cells</u> to <u>produce IL-4</u>, -5, and -13 [7].

#### Biological context of TNFSF4

- This study suggests a possible function of OX40L / OX40, through T cell-T cell interaction, in the reactivation of memory T cells in an autocrine manner, with implications for the pathogenesis of viral infections and neoplasms [2].
- These results indicated that rat OX40L can provide an efficient costimulation for rat <u>T cells</u> and that it may be involved in HTLV-1-associated <u>pathologies</u> in the rat system as has been suggested in the human system [8].
- Enhancing the immunostimulatory function of <u>dendritic cells</u> by <u>transfection</u> with mRNA encoding OX40 ligand [9].
- <u>Cell adhesion</u> assay was performed and in at least three cases, fresh ATL cells exhibited adhesion to human <u>umbilical vein endothelial cells</u> that could be considerably inhibited by either anti-OX40 MoAb or anti-gp34 MoAb [4].
- T cell proliferation by direct cross-talk between OX40 ligand on human mast cells and OX40 on human T cells: comparison of gene expression profiles between human tonsillar and lung-cultured mast cells [10].







## LLMs can be useful if used with caution

What is known about the functionality of human TNFSF4?

Human TNFSF4, also known as OX40 ligand (OX40L) or CD252, is a member of the tumor necrosis factor (TNF) superfamily. It is a costimulatory molecule that plays a critical role in the immune system by influencing T-cell activation, survival, and differentiation. Here are the key details about its functionality:

#### Structure and Expression

- TNFSF4 is expressed as a type II transmembrane protein on activated antigen-presenting cells (APCs), such as dendritic cells, B cells, and macrophages.
- It interacts with its receptor, OX40 (CD134), which is expressed on activated T cells, particularly CD4+ T cells.

# **ChatGPT**

#### 1. T-cell Costimulation

- TNFSF4 acts as a costimulatory signal during T-cell activation, enhancing the proliferation, survival, and function of T cells.
- It amplifies the immune response by promoting cytokine production, such as IL-2, which is crucial for T-cell growth.

#### 2. T-cell Memory Development

 TNFSF4-OX40 signaling supports the development and maintenance of memory T cells, ensuring a rapid and robust immune response upon re-exposure to the same antigen.

#### 3. Effector T-cell Function

 This pathway enhances the function of effector T cells, including their ability to produce cytokines like IFN-γ and TNF-α, which are essential for combating infections and tumors.

#### 4. Regulatory T-cell (Treg) Modulation

 TNFSF4 can influence Treg function, sometimes promoting their suppressive activity to prevent excessive immune responses.

#### 5. Role in Immune Diseases

- Dysregulated TNFSF4 signaling is implicated in autoimmune diseases (e.g., systemic lupus erythematosus, rheumatoid arthritis) due to its role in sustaining chronic immune activation.
- · It may also contribute to allergic diseases and inflammation.

#### 6. Role in Cancer Immunity

TNFSF4 expression on APCs can enhance anti-tumor immunity by supporting T-cell-mediated immune responses against tumors.

## Functional analysis relates hits to existing knowledge

#### Germ-line stem cell division

The self-renewing division of a germline stem cell to produce a daughter stem cell and a daughter germ cell, which will divide to form the gametes.

#### Gene/product Gene/product name

Hoxc4	homeobox C4
Ing2	inhibitor of growth family, member 2
Stra8	stimulated by retinoic acid gene 8
Zbtb16	zinc finger and BTB domain containing 16
Etv5	ets variant 5

### Advantages:

- Biological insight
- Validation of experiment
- Generate new hypotheses

### Limitations:

- You can only discover what is already known
  - Novel functionality will be missing
  - Existing annotations may be incorrect
  - Many species are poorly supported

## Functionality is generally annotated on genes

- Things to think about
  - Converting hits to genes
    - Transcripts / Proteins are easy
    - Genomic positions may be possible
  - Gene nomenclature
    - Names change over time
    - Gene definitions appear / change

- Types of list
  - Categorical (hit or not a hit)
  - Ordered
  - Quantitative

### Hits

ABC1 DEF1 GHI1 JKL1

[All non hits]

### Ordered

1. DEF1

2. ABC1

3. JKL1

4. GHI1

[All non hits]

### Quant

ABC1 = 5.3

DEF1 = 2.1

GHI1 = 7.9

JKL1 = 1.0

MNO1 = 0.4

PQR1 = 5.7

STU1 = 3.8

## Comparing your hits to functional gene sets

### Germ-line stem cell division

The self-renewing division of a germline stem cell to produce a daughter stem cell and a daughter germ cell, which will divide to form the gametes.

#### Gene/product Gene/product name

Hoxc4	homeobox C4
Ing2	inhibitor of growth family, member 2
Stra8	stimulated by retinoic acid gene 8
Zbtb16	zinc finger and BTB domain containing 16
Etv5	ets variant 5

A4galt

Atl1

Cdk19

Cdon

Cecr2

Etv5

My Hits

Flywch1

Gnpda2

Hoxc4

Ing2

ligp1

Map3k9

Mypop

Rnf6

Serinc1

Stra8

Trp73

Zbtb16

## Nothing is ever straight forward...

Best hit: "DNA Methylation" p<2e-10

name: DNA methylation

datasource: reactome

• organism: Human

idtype: hgnc symbol

• Genes:

- Methyltransferases: DNMT1 DNMT3A DNMT3B DNMT3L
- Methyltransferase targeting protein: UHRF1
- Histones!!! H2AFB1 H2AFJ H2AFV H2AFX H2AFZ H2BFS H3F3A H3F3B HIST1H2AB HIST1H2AC HIST1H2AD HIST1H2AE HIST1H2AJ HIST1H2BA HIST1H2BB HIST1H2BC HIST1H2BD HIST1H2BE HIST1H2BF HIST1H2BG HIST1H2BH HIST1H2BI HIST1H2BJ HIST1H2BK HIST1H2BL HIST1H2BM HIST1H2BN HIST1H2BO HIST1H3A HIST1H3B HIST1H3C HIST1H3D HIST1H3E HIST1H3F HIST1H3G HIST1H3H HIST1H3I HIST1H3J HIST1H4A HIST1H4B HIST1H4C HIST1H4D HIST1H4E HIST1H4F HIST1H4H HIST1H4I HIST1H4J HIST1H4K HIST1H4L HIST2H2AA3 HIST2H2AA4 HIST2H2AC HIST2H2BE HIST2H3A HIST2H3C HIST2H3D HIST2H4A HIST2H4B HIST3H2BB HIST4H4

## Sources of functional gene lists

- Human curated
  - Gene Ontology
  - Biological Pathways
- Domains / Patterns
  - Protein functional domains
  - Transcription factor regulated
- Experimental
  - Co-expressed genes
  - Interactions
  - Hits from other studies













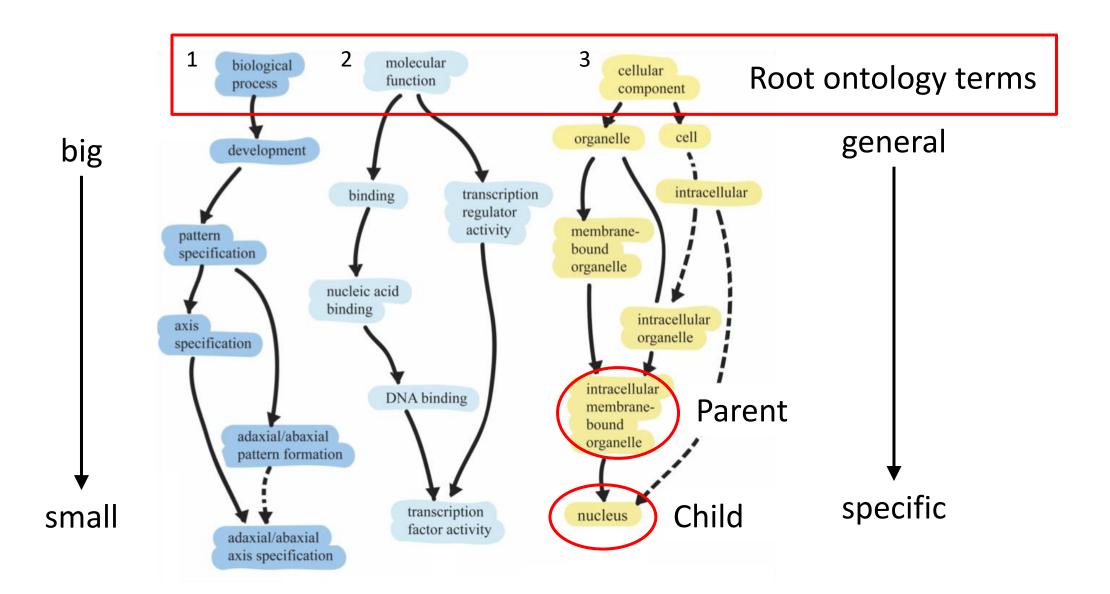


## Gene Ontology is a human curated functional database





## GO has three domains and a hierarchical structure



## Genes are specifically placed into each domain

## Nanog homeobox

#### Cellular Component

GO:0005634 nucleus

GO:0005654 nucleoplasm

GO:0005730 nucleolus

#### Molecular Function

GO:0003677 DNA binding

GO:0003700 transcription factor activity

GO:0003714 transcription corepressor activity

GO:0005515 protein binding

GO:0043565 sequence-specific DNA binding

#### Biological Process

GO:0001714 endodermal cell fate specification

GO:0006351 transcription, DNA-templated

GO:0006355 regulation of transcription, DNA-templated

GO:0007275 multicellular organism development

GO:0008283 cell proliferation

GO:0019827 stem cell population maintenance

GO:0030154 cell differentiation

GO:0035019 somatic stem cell population maintenance

GO:0045595 regulation of cell differentiation

GO:0045944 positive regulation of transcription from RNA pol2

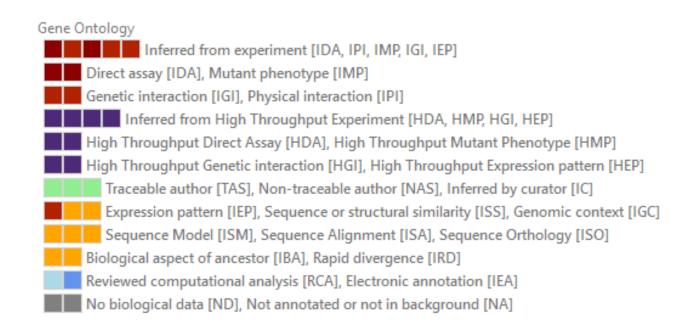
GO:1903507 negative regulation of nucleic acid-templated

transcription

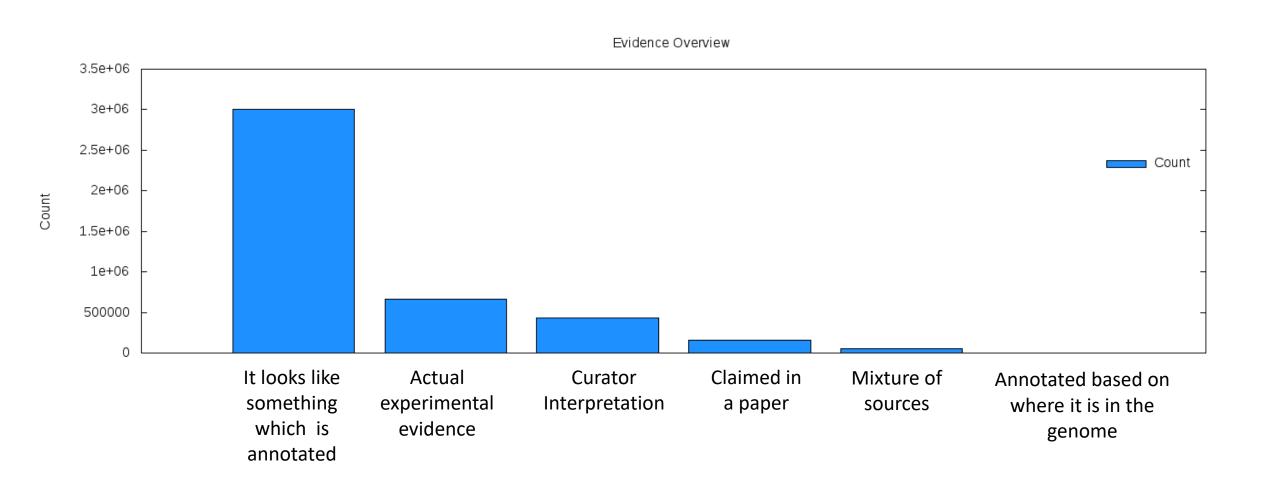
### GO Annotations come with evidence

- Experimental
  - Experiment (EXP)
  - Direct Assay (IDA)
  - Physical Interaction (IPI)
  - Mutant Phenotype (IMP)
- Computational
  - Sequence Similarity (ISS)
  - Sequence Model (ISM)
  - Genomic Context (IGC)
  - Biological aspect of Ancestor (IBA)
  - Key Residues (IKR)

- Publications
- Curators



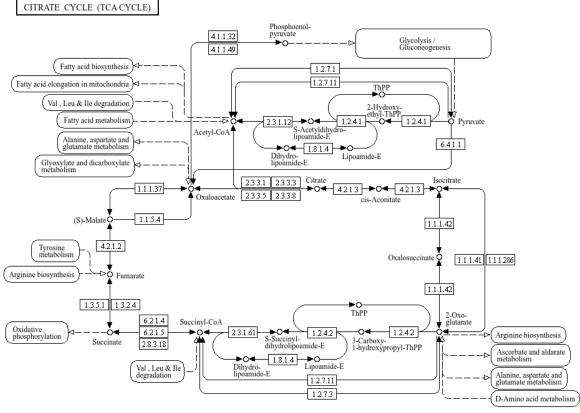
### Annotations come with evidence



# Pathway databases trace metabolic pathways and their regulation





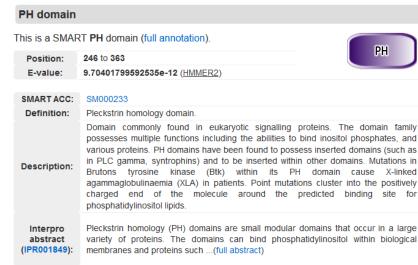


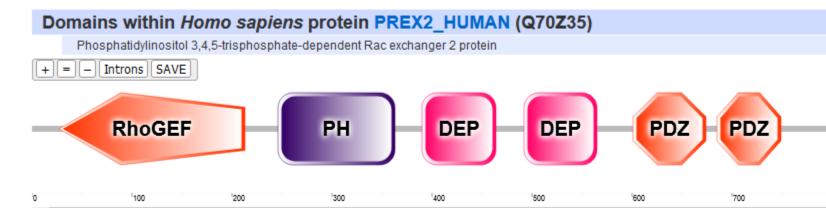


# Protein Domain databases annotate functional subdomains within proteins

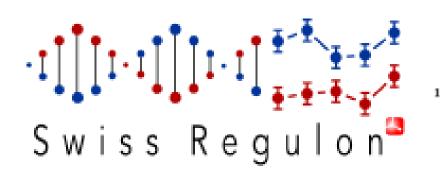




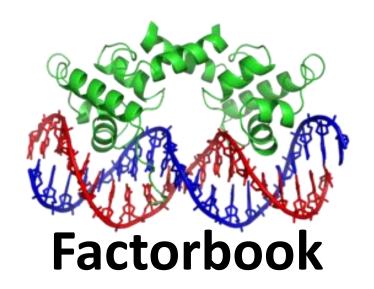




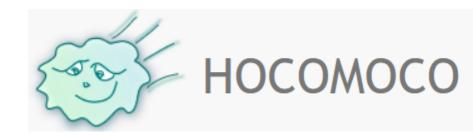
# Transcription Factor databases group genes by the motifs in their promoters







AnimalTFDB v4.0



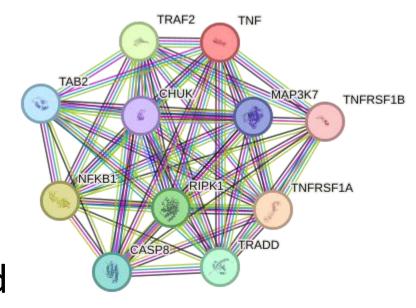
# Interaction databases map out interactions between genes / proteins







- Physical interaction
- Genetic interaction
- Gene fusions
- Literature mentions
- Genome neighbourhood

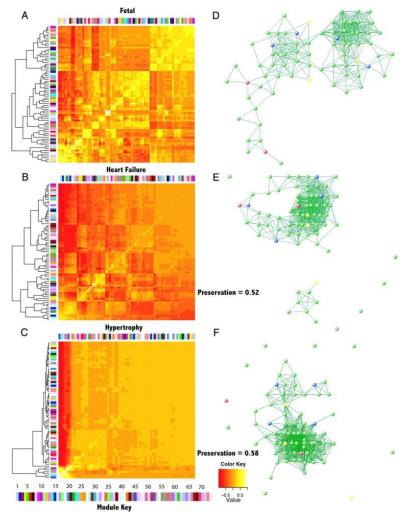


# Co-expression databases group genes which are expressed together











#### mouse-ortholog hallmark gene

MSigDB Hallmoods a little MSigDB Hallmarks collection mapped to their mouse orthologs.

positional gene sets corresponding to mouse chromosome cytogenetic bands.

curated gene sets from online pathway databases, publications in PubMed, and knowledge of domain experts.

regulatory target gene sets based on M3 gene target predictions for microRNA seed sequences and predicted transcription factor binding sites.

M5 ontology gene sets consist of genes

cell type signature gene sets curated M8 from cluster markers identified in singlecell sequencing studies of mouse tissue.

hallmark gene sets are coherently expressed signatures derived by aggregating many MSigDB gene sets to represent well-defined biological states or processes.

ontology gene sets consist of genes annotated by the same ontology term.

positional gene sets corresponding to C1 human chromosome cytogenetic bands.

oncogenic signature gene sets defined C6 directly from microarray gene expression data from cancer gene perturbations.

curated gene sets from online pathway C2 databases, publications in PubMed, and knowledge of domain experts.

immunologic signature gene sets represent cell states and perturbations within the immune system.

regulatory target gene sets based on gene target predictions for microRNA seed sequences and predicted transcription factor binding sites.

cell type signature gene sets curated C8 from cluster markers identified in single-cell sequencing studies of human tissue.

computational gene sets defined by Mining large collections of cancer-oriented expression data.

https://www.gsea-msigdb.org/gsea/msigdb

# Testing for enriched gene sets



## There are two basic ways to test for enrichment

### Categorical

- Start from a list of hit genes
- Count overlaps between hit list and functional list
- Find Functional lists where the degree of overlap is statistically unlikely

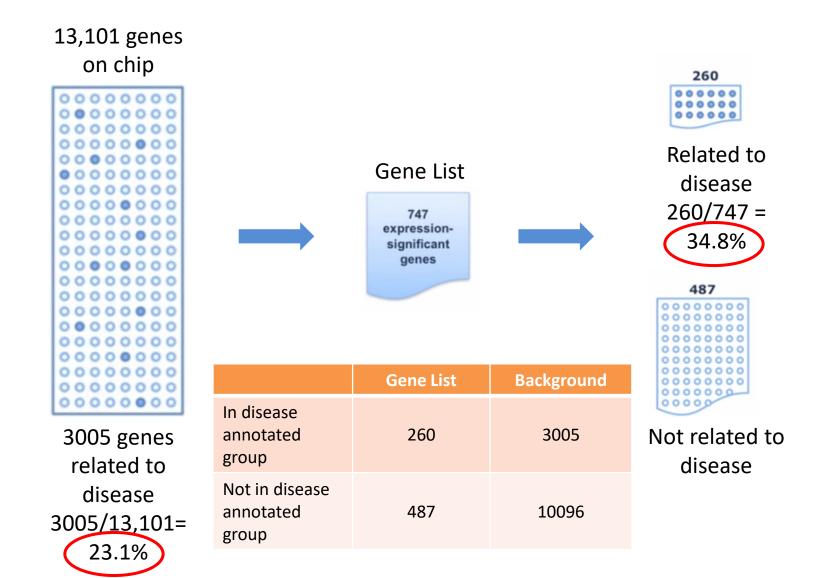
### Quantitative

- Start with all genes
- Associate a value with each gene
- Look for functional sets with unusual distributions of values

## Categorical Enrichment Analysis



## Categorical tests for enrichment



## Fisher's Exact test

	Gene List	Background	Total
In disease annotated group	260 E = 176.1	3005 E = 3088.8	3265
Not in disease annotated group	487 E = 570.9	10096 E = 10012.1	10583
Total	747	13101	13848

```
> counts <-(matrix(data = c(260, 487, 3005, 10096), nrow = 2))
> fisher.test(counts)

Fisher's Exact Test for Count Data

data: counts
p-value = 9.769e-13
alternative hypothesis: true odds ratio is not equal to 1
95 percent confidence interval:
1.52846 2.10120
sample estimates:
odds ratio
1.793564

(260/487) / (3005/10096)
```

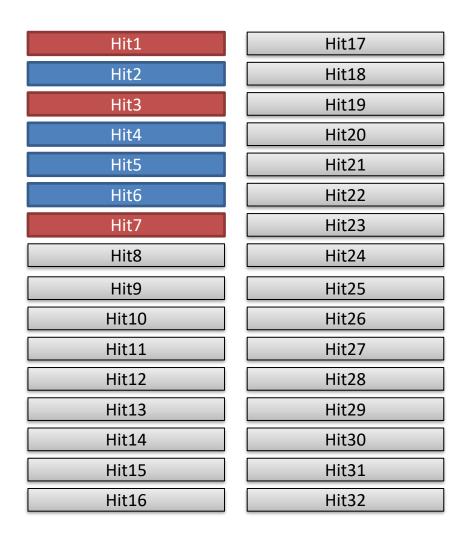
# Categorical tests are influenced by where you set the cutoff for "interesting" genes

Hit17
Hit18
Hit19
Hit20
Hit21
Hit22
Hit23
Hit24
Hit25
Hit26
Hit27
Hit28
Hit29
Hit30
Hit31
Hit32

#### Function X

- 3 hits out of 32 in 'interesting' list
- Not significant(p=0.07)

# Categorical tests are influenced by where you set the cutoff for "interesting" genes



### Function X

- 3 hits out of 7 in 'interesting' list
- Significant (p=0.02)

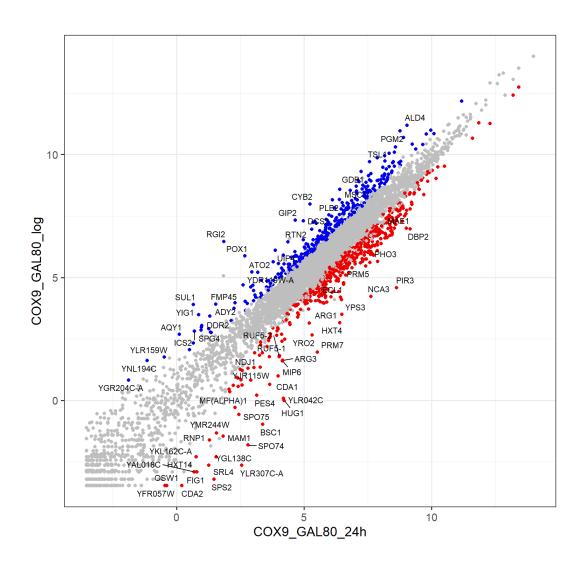
# Ordered, but not quantitative lists allow sequential categorical analysis

Hit1	Hit17
Hit2	Hit18
Hit3	Hit19
Hit4	Hit20
Hit5	Hit21
Hit6	Hit22
Hit7	Hit23
Hit8	Hit24
Hit9	Hit25
Hit10	Hit26
Hit11	Hit27
Hit12	Hit28
Hit13	Hit29
Hit14	Hit30
Hit15	Hit31
Hit16	Hit32

#### Function X

- Length=1 p=0.60
- Length=2 p=0.80
- Length=3 p=0.30
- Length=4 p=0.35
- Length=5 p=0.40
- Length=6 p=0.45
- Length=7 p=0.05
- Length=8 p=0.08
- Length=9 p=0.10

## **Directional Gene Lists**



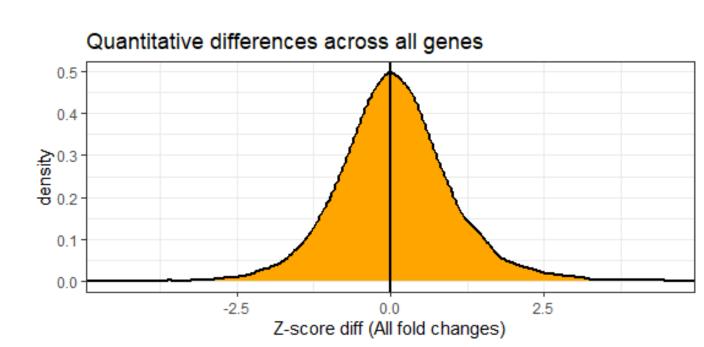
- One search or two?
  - One search
    - Higher power (more genes)
    - Lower enrichment
    - Mixed effects (pathways)
  - Two searches
    - Easier interpretation
    - Less power
    - Higher enrichment

## Quantitative Enrichment Analysis

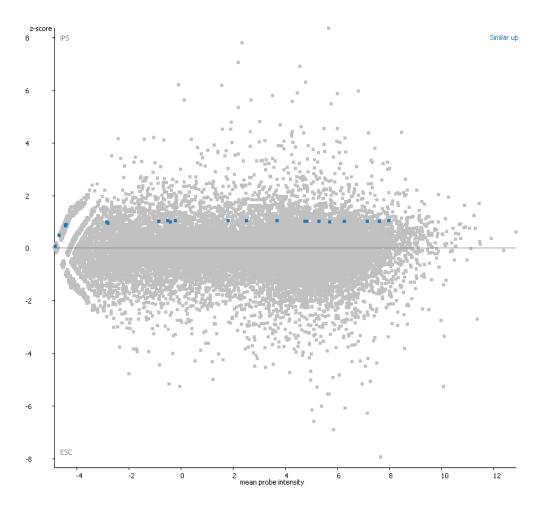


## Quantitative comparisons can offer more power

- What quantitative value can we use?
  - Differential p-value (normally -10 log(p))
  - Fold change
  - Absolute difference



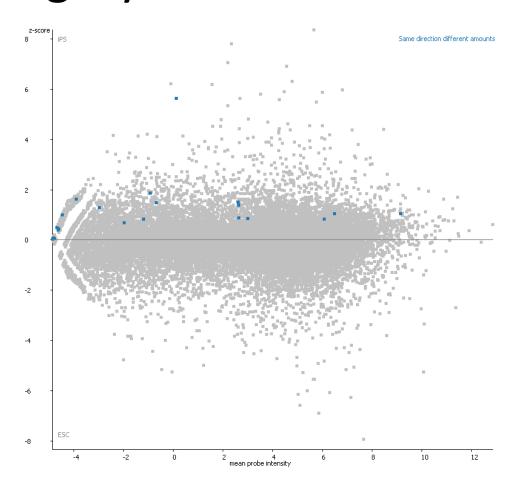
# What kind of changes do we expect in an interesting category?



Student's T-test

Genes in that category all change, and by about the same amount?

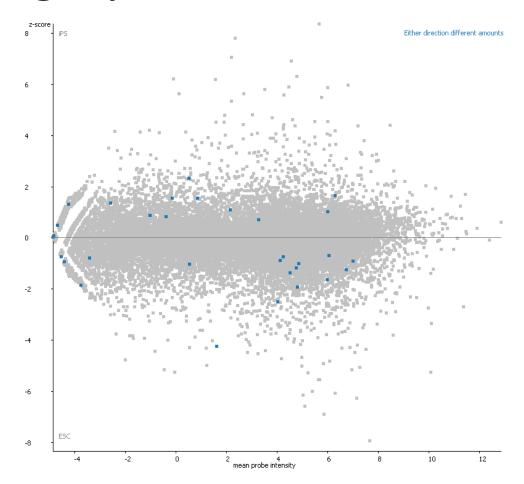
# What kind of changes do we expect in an interesting category?



Kolmogorov Smirnov Test

Genes in that category all change in the same direction, but by different amounts?

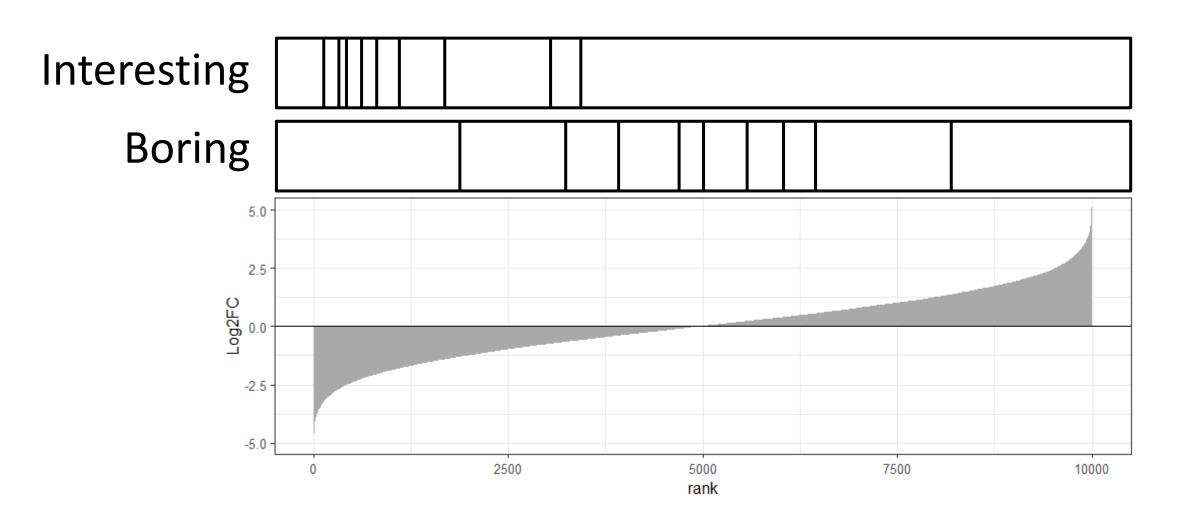
# What kind of changes do we expect in an interesting category?



Absolute KS Test

Genes in that category all change in either direction, but by different amounts?

#### **GSEA** statistics

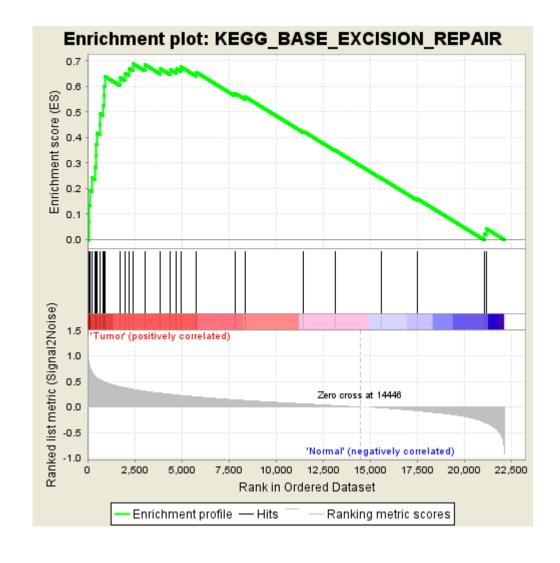


#### **GSEA Statistics**

- Keep a running total
- Start at the highest values
- If gene is in the set add value
- Otherwise subtract value

Enrichment score is max score

Stats compare ES with randomly shuffled data



## Multiple Testing Correction

- Original p-value is for one test (one gene set)
- Thousands of sets tested in each analysis
- Many tools report raw as well as corrected p-values

#### **FALSE POSITIVES**

Raw p-values (no correction)

Benjamini & Hochberg False Discovery Rate (FDR)

Bonferroni Corrected P-value

## What do we get back from an enrichment test?

#### A p-value

 Remember that this reflects not only difference but also variance and power (number of observations)

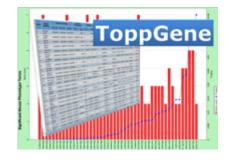
#### A difference value

- Enrichment difference (odds ratio)
- Mean quantitative difference
- Remember large differences are easier to obtain with small numbers of observations

#### Tools for functional gene list analysis

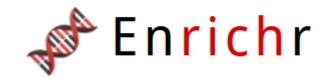
 There are many different tools available, both free and commercial

Popular tools include:

















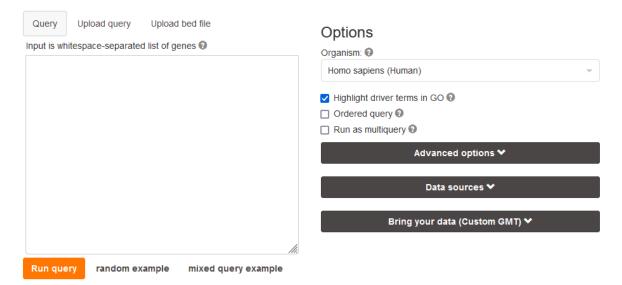






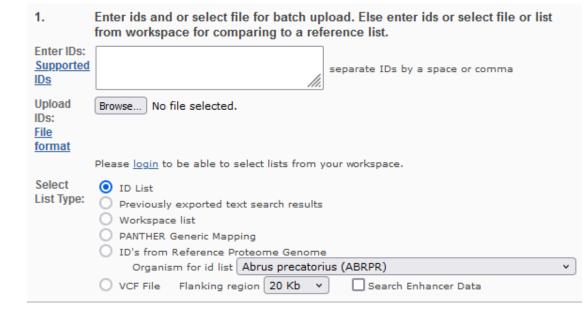


- Categorical or ordered statistics
- Lots of additional options
- Wide species support
- Interesting presentation
  - Doesn't scale well to lots of hits





- Categorical or Quantitative statistics
- Part of Gene Ontology Consortium
  - Annotations are up to date
- Simple enrichment analysis
- Functional lists and categorical break down







#### Your data

**Options** 

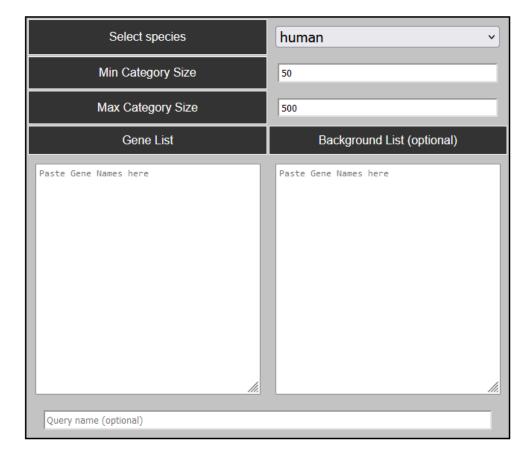
**Analysis** 

Step 1: Select a file from your computer or paste your own data and click on the corresponding "Continue" button.

- Categorical or quantitative statistics
- Pathway focussed
- Simple submission interface (no custom background)
- Really nice visualisations

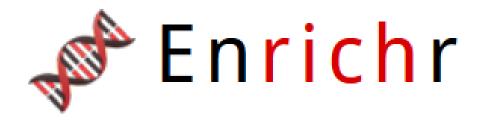
# GOliath

- Categorical statistics
- Limited species support
- Allows custom backgrounds
- Uses PathwayCommons gene sets
- Innovative detection and presentation of artefacts





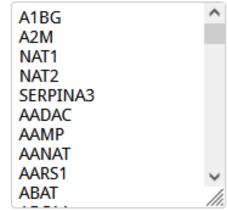
- Categorical Statistics
- Most popular system (mostly historic)
- Has been behind the latest annotation
  - Was updated again, but now behind once more
- Lots of support for different IDs and Species
- Configurable gene sets
- Simple output presentation



- Categorical Statistics
- Biggest selection of gene sets
- Simple interface, but limited options
  - No species information
- Simple interactive visualisation
- Novel scoring scheme to rank hits

Drop a file or paste a set of Entrez gene symbols on each row in the textbox below. You can try a gene set example. Also, you can now try adding a background (clear).

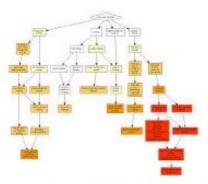




0 gene(s) entered







Gene Ontology enRIchment anaLysis and visuaLizAtion tool

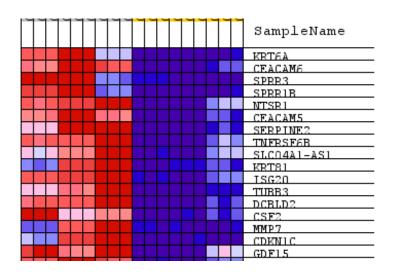
- Categorical or ranked analysis
- Mostly GO gene list support
- Interesting visualisation options

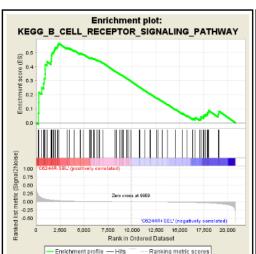
ina visuaLi_Ation tool
Step 1: Choose organism
Homo sapiens v
Step 2: Choose running mode
O Single ranked list of genes    O Two unranked lists of genes (target and background lists)
Step 3: Paste a ranked list of gene/protein names
Names should be separated by an <enter>. The preferred format is gene symbol. Other supported formats are: gene and protein RefSeq, Uniprot, Unigene and Ensembl.</enter>
Target set:
Or upload a file: Browse No file selected.
Background set:
Or upload a file: Browse No file selected.

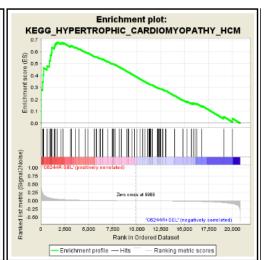
#### **GSEA**



- Quantitative enrichment
- Designed for expression datasets
- Local application
- Imports tab delimited expression data









#### SeqMonk Mapped Sequence Data Analyser

Version: 1.48.2.devel

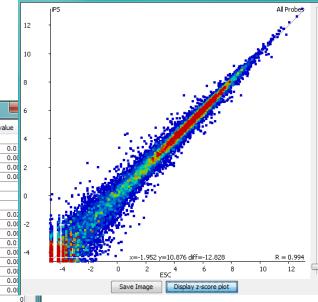
www.bioinformatics.babraham.ac.uk/projects/

© Simon Andrews, Laura Biggins Babraham Bioinformatics, 2006-21

HTSJDK BAM/SAM reader @The Broad Institute, 2009-19



Local Java application



Gene Set Results

core	1 ips	S	Term	No of p	mean z	identifier	descript	p-value
	and extension in order to provide the control of th		cell migration involved in g	13	-2.288	CELL MIG	CELL MIG	
	ceir nigration in voived in gastruiation		primitive streak formation	10	-1.3	PRIMITIV	PRIMITIV	0.0
			2'-5'-oligoadenylate synth	10	-1.258	2'-5'-OLI	2'-5'-OLI	0.00
			cytoplasmic dynein complex	26			CYTOPLA	0.00
	A Common		host cell part	15			HOST CE	0.00
	1		semaphorin-plexin signalin	27		SEMAPH		
	10 m		cellular response to interfe	36			CELLULA	
	102.00 (Control of Control of Con		response to interferon-beta	43			RESPONS	
			fascia adherens	12			FASCIA	0.02
			generation of neurons	14			GENERAT	0.00
			low-density lipoprotein rec	10		LOW-DE		0.00
			collagen-activated tyrosin	10			COLLAGE	0.0
			smooth muscle cell migration	10			SMOOTH	0.00
			antigen processing and pr	11			ANTIGEN	0.00
			inactivation of mapk activity	14			INACTIV	0.00
		H	positive regulation of macr	11			POSITIVE	0.00
		H	signal transduction involve profilin binding	23 10				0.00
		H	response to interferon-alpha	19		PROFILI	RESPONS	0.00
		H	fibrillar collagen trimer	19			FIBRILLA	0.00
		H	negative regulation of acti	11			NEGATIV	0.00
		H	limb bud formation	10		LIMB BU		0.00
			regulation of glial cell migr	11			REGULAT	0.00
			semaphorin receptor activity	11		SEMAPH		0.00
	ESC		left/right axis specification	11			LEFT/RIG	0.00
	x=-5.474 y=7.148	H	collagen-activated signalin	13			COLLAGE	0.01
	-4 -2 0 2 4 6 8 10 12	H	cellular response to choles	10			CELLULA	0.05
	-4 -2 U Z 4 6 8 10 12 mean probe intensity	H	enithelial cell differentiatio	16		FPITHFI I		0.0.

# Gene List Practical

https://tinyurl.com/exercisetostartat



# **Exploring and Presenting** Results

Simon Andrews, Laura Biggins

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#### Functional enrichment results

- Gene set information
  - Gene set name
  - Gene set source
  - Gene set description

- Statistical information
  - Raw p-value
  - Corrected p-value
  - Enrichment value

- Count information
  - Hit genes in category
  - Hit genes outside category
  - Background genes in category
  - Background genes outside category

#### Functional enrichment results

- Gene set information
  - Gene set name
  - Gene set source
  - Gene set description

- Statistical information
  - Raw p-value
  - Corrected p-value
  - Enrichment value

- Count information
  - Hit genes in category
  - Hit genes outside category
  - Background genes in category
  - Background genes outside category

# Tables are often enough

Category	<i>P</i> value	Genes in GO category over-expressed	% of differentially expressed genes in GO category	Genes in GO category on array	% genes on array ir GO category
Over-expressed in AJC: Biological process					
GO:9058: biosynthesis	0.009	52	24.41	1264	17.82
GO:7610: behavior	0.019	10	4.695	156	2.2
Over-expressed in AJC: Molecular function					
GO:5198: structural molecule activity	< 0.001	43	17.92	750	9.043
Over-expressed in SL: Biological process					
GO:8152: metabolism	0.019	192	71.91	4674	65.91
Over-expressed in SL: Molecular function					
GO:16209: antioxidant activity	0.013	6	1.917	52	0.627
GO:8135: translation factor activity, nucleic acid binding	0.010	13	4.153	166	2.001
GO:45182: translation regulator activity	0.014	13	4.153	173	2.086
GO:5489: electron transporter activity	0.005	17	5.431	225	2.713
GO:8233: peptidase activity	0.043	28	8.946	529	6.378
GO:3824: catalytic activity	0.001	166	53.04	3645	43.95

These are the significant GO Slim categories representing both biological process and molecular function ontologies for population specific significantly over-expressed ( $P \le 0.05$ ; no multiple test correction) features. For each significant GO category, we include the P value number of over-expressed genes in that GO, percentage of representation in the over-expressed list, number of features of that GO in the microarray, and percentage of representation on the entire microarray.

	Gene Ontology Term	% <sup>1</sup>	Univariate p-value <sup>2</sup>	FDR-adjusted p-value <sup>3</sup>				
	Immunoglobulin		4.6E-25	1.8E-23				
D/D	Immunoglobulin V-set		2.6E-18	2.5E-17				
B/P Cluster	Antigen binding		8.7E-16	1.2E-14				
Olusio	Immunoglobulin-like fold		9.7E-16	4.8E-15				
	Immune response	41.4	2.0E-13	2.6E-11				
			. ==					
	Positive regulation of immune system process	24.4	1.7E-08	2.5E-05				
T/NU/	Natural killer cell mediated cytotoxicity	19.5	9.7E-07	5.9E-05				
T/NK Cluster	Positive regulation of lymphocyte activation	17.1	3.3E-07	6.9E-05				
Olusiei	T-cell	12.2	1.3E-06	7.2E-05				
	Positive regulation of lymphocyte differentiation	12.2	3.7E-06	3.3E-04				
	MHC class II, alpha/beta chain, N-terminal	39.1	7.0E-22	3.4E-20				
M/D	Class II histocompatibility antigen	39.1	1.3E-19	1.9E-18				
M/D Cluster	MHC class II protein complex	39.1	4.6E-20	3.3E-18				
3140101	Immunoglobulin C1-set	43.5	7.0E-18	1.7E-16				
	Antigen processing and presentation	47.8	1.3E-18	3.0E-16				
Tporconto	are of cluster genes (relative to all genes on array) appointed for a given entelogy term:							

<sup>&</sup>lt;sup>1</sup> percentage of cluster genes (relative to all genes on array) annotated for a given ontology term; <sup>2</sup> modified Fisher's Exact Test; <sup>3</sup> Benjamini and Hochberg false discovery rate-adjusted *p* value

## **Graphical Representations**

- Need to add something over a table
  - Relationships between multiple result values
  - Representation of redundancy between categories
  - Relationship to original data
  - Context of surrounding pathway

## Plotting relationships between values

DNA damage checkpoint DNA integrity checkpoint Mitotic DNA damage checkpoint

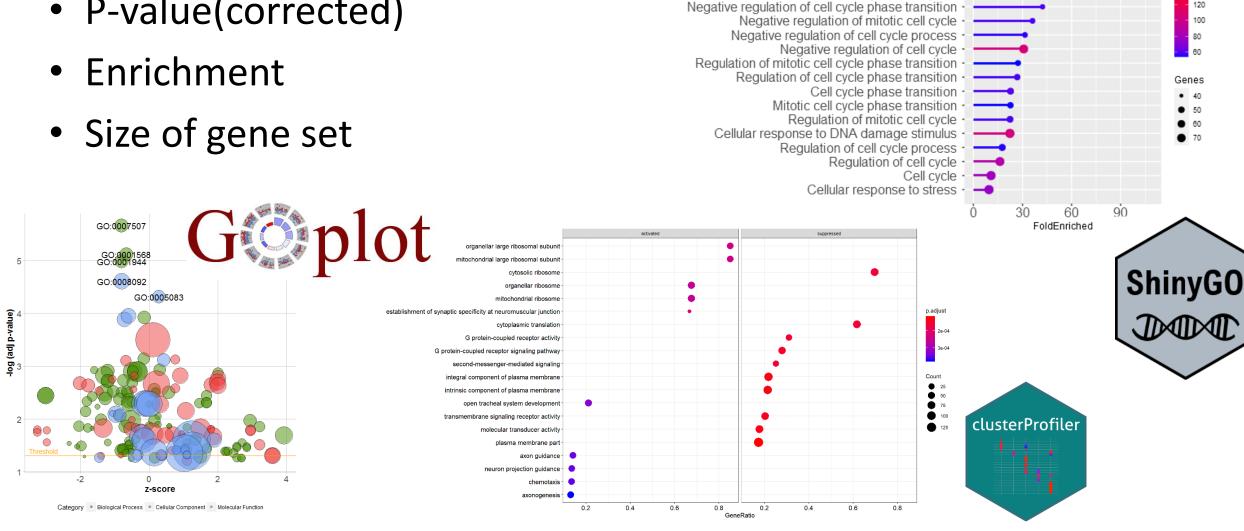
Mitotic DNA integrity checkpoint

Mitotic cell cycle checkpoint

Cell cycle checkpoint

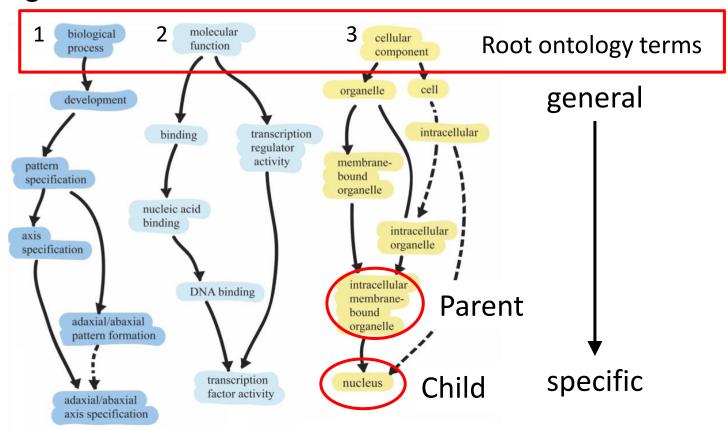
EnrichmentFDR

P-value(corrected)



## Redundancy in gene lists

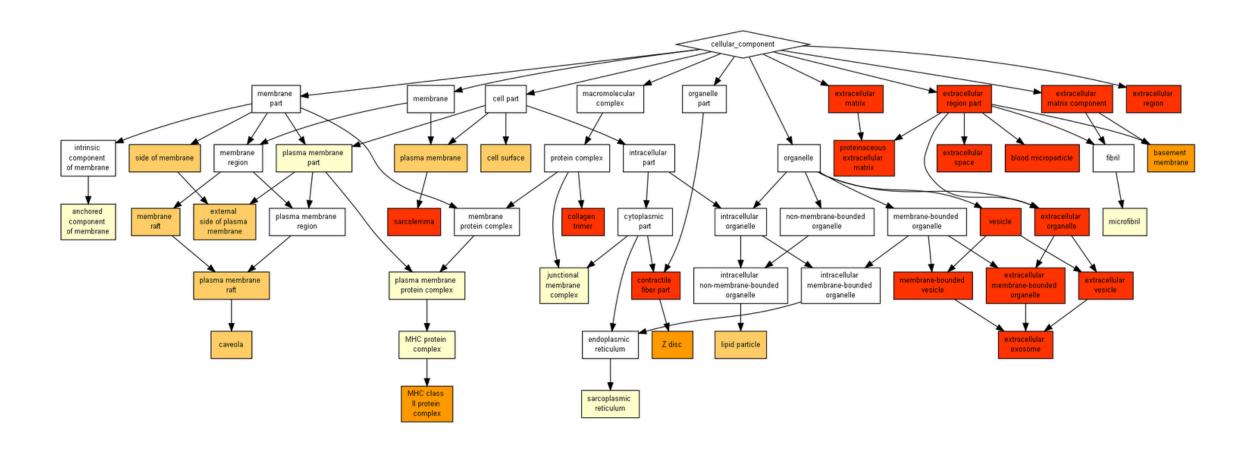
Gene ontology is hierarchical - a gene is placed in the most specific category and will also appear in all the parent categories



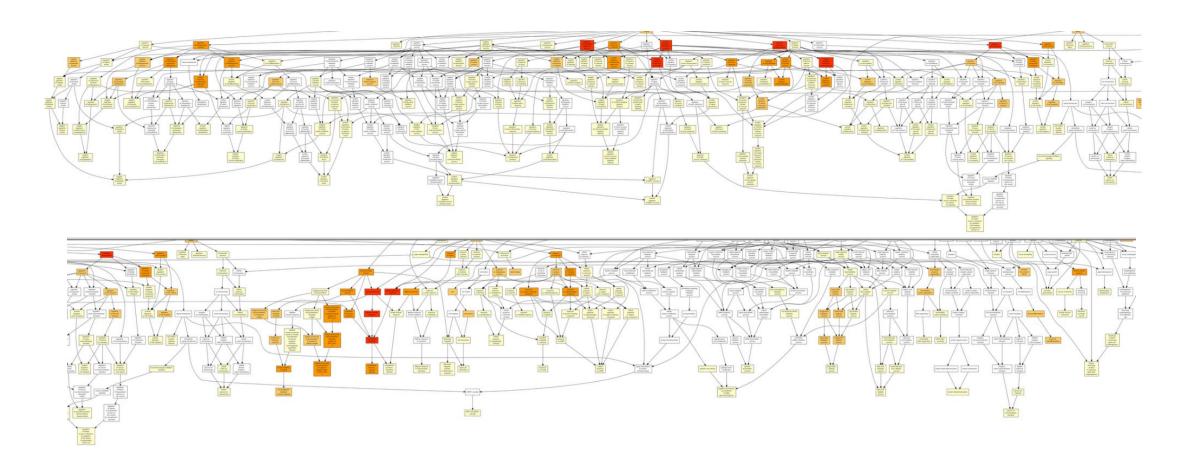
## Redundancy: DAVID clustering

Annotation Cluster 1				40 Cluster(s) Download File									
	Enrichment Score: 16.36	G	<b></b>	Count	P_Value	Benjamini							
SP_PIR_KEYWORDS	<u>dna-binding</u>	<u>RT</u>		53	3.5E-24 4	4.5E-22							
GOTERM_BP_FAT	regulation of transcription	<u>RT</u>		60	2.0E-20	1.8E-17							
GOTERM_MF_FAT	DNA binding	<u>RT</u>		54	6.0E-20 7	7.9E-18							
GOTERM_MF_FAT	transcription regulator activity	<u>RT</u>		45	2.0E-19	1.3E-17							
SP_PIR_KEYWORDS	transcription regulation	<u>RT</u>		49	5.9E-19 3	3.8E-17							
GOTERM_MF_FAT	sequence-specific DNA binding	<u>RT</u>	_	30	1.5E-16 4	4.9E-15							
SP_PIR_KEYWORDS	Transcription	<u>RT</u>		48	8.1E-16	3.3E-14							
GOTERM_BP_FAT	transcription	<u>RT</u>		48	1.9E-15 8	8.1E-13							
GOTERM_MF_FAT	transcription factor activity	<u>RT</u>	_	33	2.8E-15 9	9.1E-14							
SP_PIR_KEYWORDS	nucleus	<u>RT</u>		69	1.1E-14 3	3.6E-13							
GOTERM_BP_FAT	regulation of RNA metabolic process	<u>RT</u>	_	40	2.1E-12 6	6.1E-10							
GOTERM_BP_FAT	regulation of transcription, DNA-dependent	<u>RT</u>	_	39	6.4E-12	1.4E-9							
Annotation Cluster 2	Enrichment Score: 10.03	G	<u>™</u>	Count	P_Value	Benjamini							
GOTERM_MF_FAT	sequence-specific DNA binding	<u>RT</u>	_	30	1.5E-16 4	4.9E-15							
GOTERM_MF_FAT	transcription factor activity	<u>RT</u>	=	33	2.8E-15 9	9.1E-14							
INTERPRO	<u>Homeodomain-related</u>	<u>RT</u>	=	16	2.3E-10 4	4.3E-8							
INTERPRO	<u>Homeobox</u>	<u>RT</u>	=	15	1.8E-9	1.7E-7							
INTERPRO	Homeobox, conserved site	<u>RT</u>	=	14	3.4E-9	2.1E-7							
SP_PIR_KEYWORDS	Homeobox	<u>RT</u>		15	8.5E-9	1.8E-7							
UP_SEQ_FEATURE	DNA-binding region:Homeobox	<u>RT</u>	=	13	2.6E-8	3.7E-6							
SMART	HOX	<u>RT</u>	=	15	4.7E-8	2.1E-6							
Annotation Cluster 3	Enrichment Score: 5.86	G	<u>™</u>	Count	P_Value	Benjamini							
INTERPRO	<u>Transcription factor, fork head, conserved</u> <u>site</u>	<u>RT</u>	=	7	3.6E-7	1.7E-5							
INTERPRO	Transcription factor, fork head	<u>RT</u>	=	7	3.6E-7	1.7E-5							
UP_SEQ_FEATURE	DNA-binding region:Fork-head	<u>RT</u>	=	7	9.1E-7 6	6.5E-5							
SMART	<u>FH</u>	<u>RT</u>	=	7	1.8E-6	4.0E-5							
INTERPRO	Winged helix repressor DNA-binding	<u>RT</u>	=	9	2.5E-5	6.6E-4							

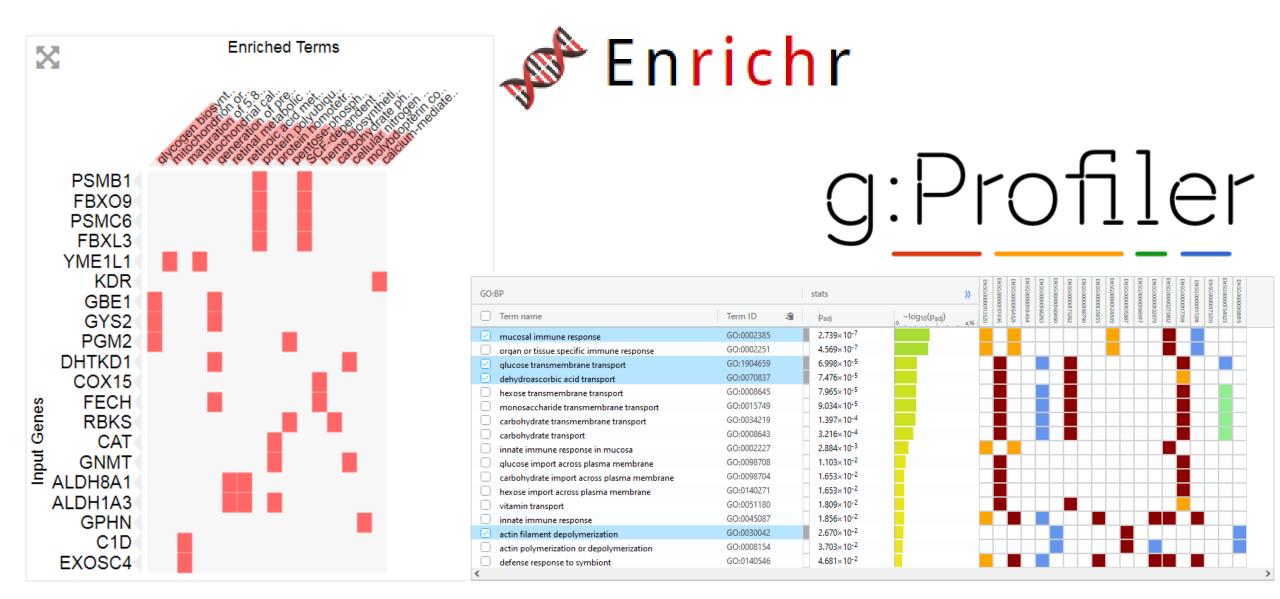
## Redundancy: Gorilla GO images



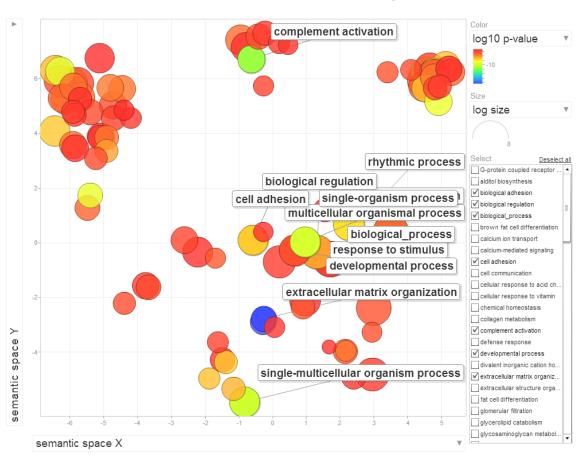
## Redundancy: Gorilla GO images



#### Redundancy: Overlap plots



## 2D Redundancy

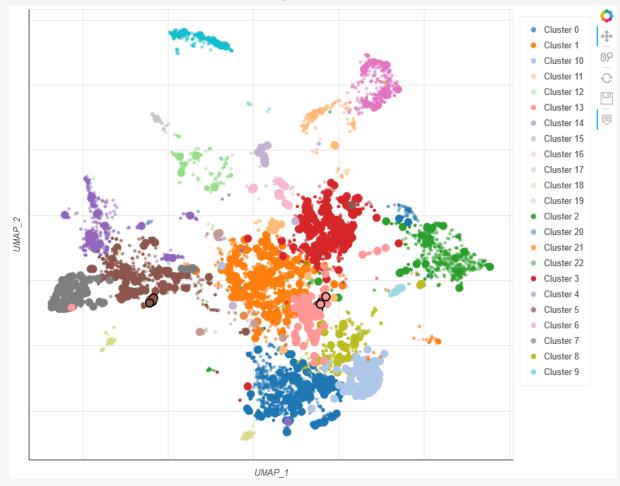


Revigo (from Gorilla) http://revigo.irb.hr/

# Enrichment Analysis Visualisation (from Enrichr)

https://appyters.maayanlab.cloud/Enrichment\_Analysis\_Visualizer/

Scatter plot visualization for GO\_Biological\_Process\_2023.

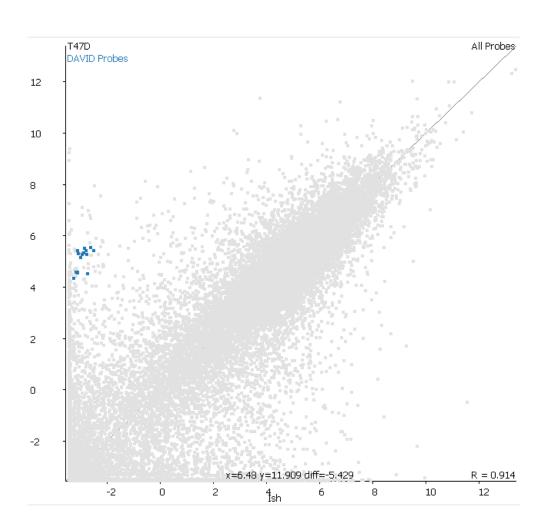


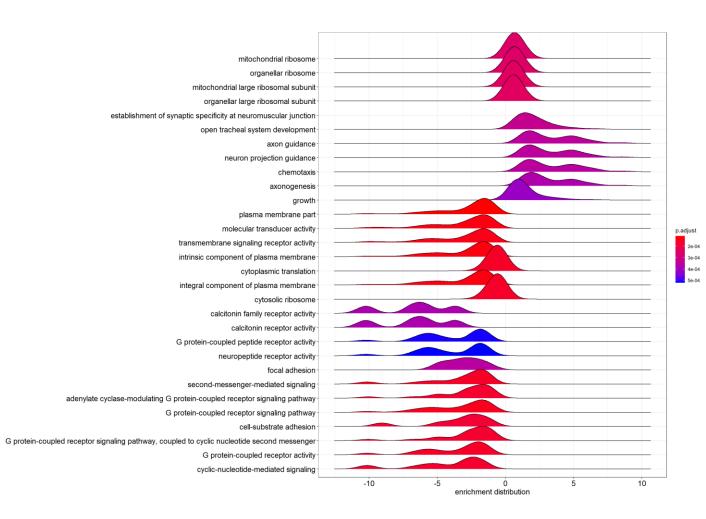
## Relationship to original data

- Quantitative values for genes in category
  - Direction and magnitude of change

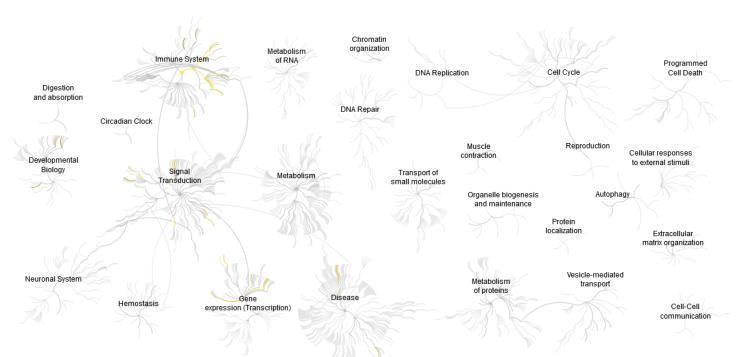
- Look at genes in category which aren't hits
  - Relative numbers
  - Supportive changes?

## Relationship to original data



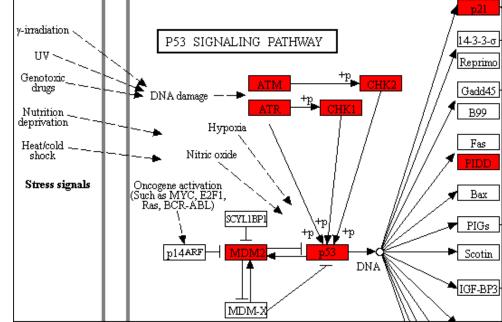


## Pathways

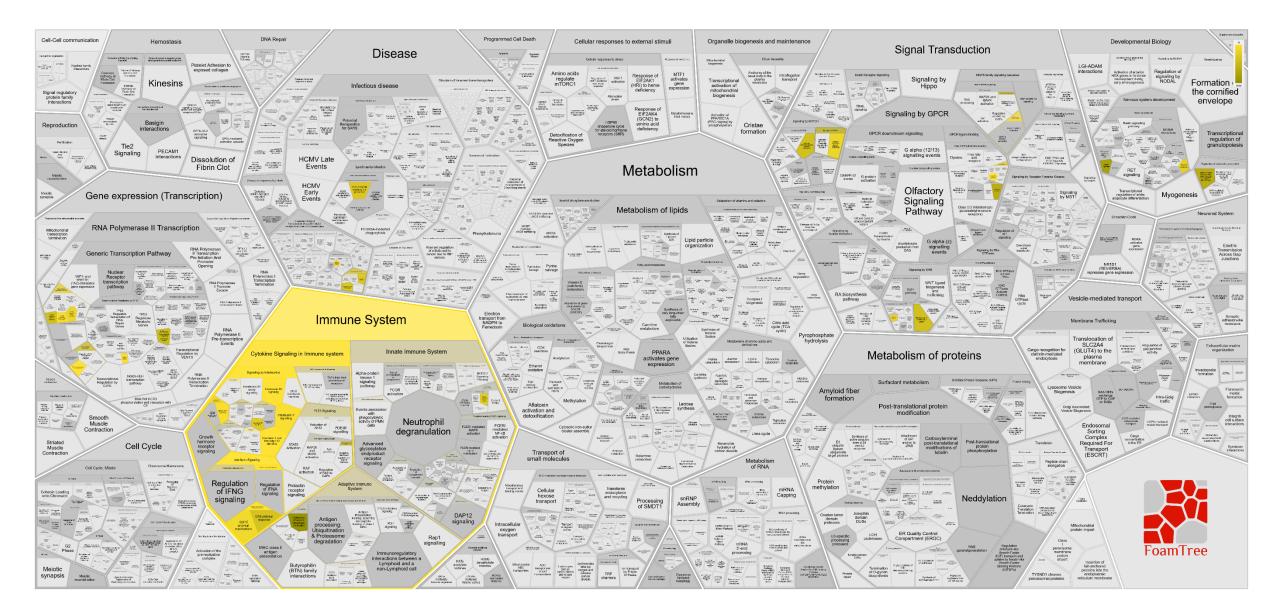


#### Reactome

#### ShinyGO



## Pathways: Reactome



#### Summary

- Tables are often sufficient
  - Must include name, enrichment, corrected p-value
  - Other values are useful, but don't put in everything
- Figures can add extra information
  - Plotting multiple metrics
  - Illustrating redundancy
  - Relating to original data
  - Mapping to pathways



# Artefacts and Biases in Gene Set Analysis

Simon Andrews, Laura Biggins, Christel Krueger

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## What does gene set enrichment test?

• Is a functional gene set enriched for genes in my hit list compared to a background set

 Are some genes more likely to turn up in the hits for technical reasons?

 Are some genes never likely to turn up in the hit list for technical reasons?

#### Biases

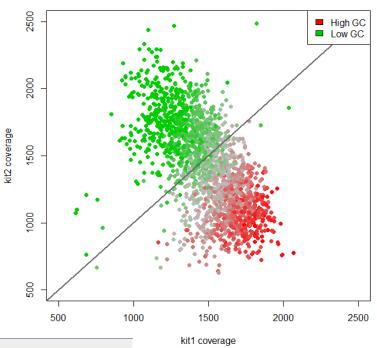
- All datasets contain biases
  - Technical
  - Biological
  - Statistical

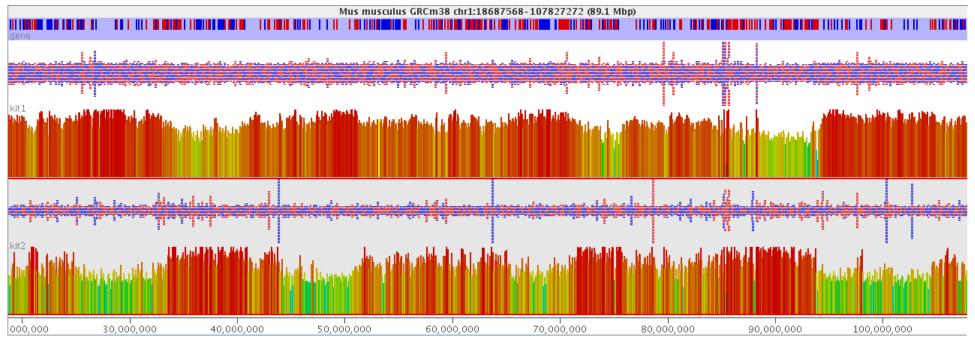
Biases can lead to incorrect conclusions

- We should be trying to spot these
  - Some are more obvious than others!

#### **Technical Biases**

Simple GC bias from different polymerases in PCR





## Statistical Biases

- The power to detect a significant effect is based on:
  - How big the change is
  - How well observed the data is (sample size)

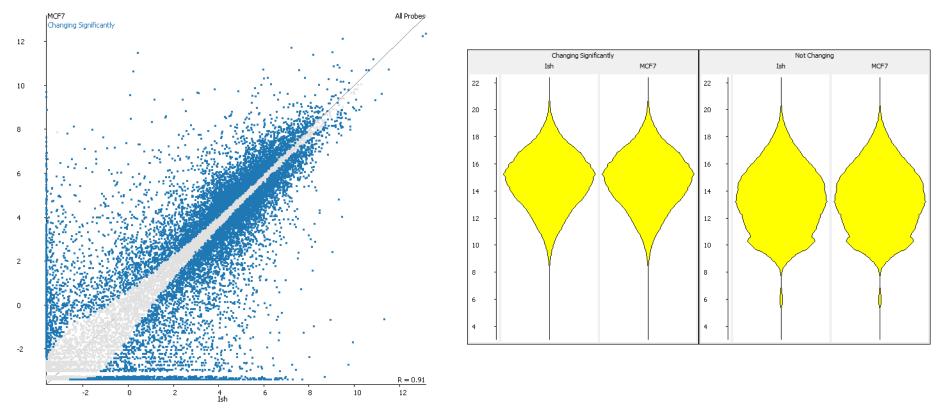
Lists of hits are often biased based on statistical power

# RNA-Seq Statistical Biases

What determines whether a gene is identified as significantly differentially regulated?

- The amount of change (fold change)
- The variability
- How well observed was it
  - How much sequencing was done overall?
  - How highly expressed was the gene?
  - How long was the gene?
  - How mappable was the gene?

# RNA-Seq Statistical Biases



- Unlikely to ever see hits from genes which are
  - Lowly expressed
  - Short

# **Biological Biases**



# Biases Look Like Real Biology

Bias	Function	P-Value
High GC	DNA-Templated Transcription	2.00E-20
Low GC	GPCR Signalling	4.00E-12
Long Genes	Synapse	2.30E-30
Chr 18	Homophilic Cell Adhesion	1.01E-26

Hindawi Publishing Corporation Bvidence-Based Complementary and Alternative Medicine Volume 2016, Article ID 7276161, 13 pages http://dx.doi.org/10.1155/2016/7276161

#### Research Article

#### Epigenetic Profiling of H3K4Me3 Reveals Herbal Medicine Jinfukang-Induced Epigenetic Alteration Is Involved in Anti-Lung Cancer Activity

Jun Lu, Xiaoli Zhang, Tingting Shen, Chao Ma, Jun Wu, Hualei Kong, Jing Tian, Zhifeng Shao, Xiaodong Zhao, 2 and Ling Xu<sup>2,4</sup>

Gene Ontology analysis indicates that these genes are involved in tumor-related pathways, including pathway in cancer, basal cell carcinoma, apoptosis, induction of programmed cell death, regulation of transcription (DNA-templated), intracellular signal transduction, and regulation of peptidase activity.

Traditional Chinese medicine Jinfukang (JFK) has been clinically used for treating lung cancer. To examine whether epigenetic modifications are involved in its anticancer activity, we performed a global profiling analysis of H3K4Me3, an epigenomic marker associated with active gene expression, in JFK-treated lung cancer cells. We identified II,670 genes with significantly altered status of H3K4Me3 modification following JFK treatment (P < 0.05). Gene Ontology analysis indicates that these genes are involved in tumor-related pathways, including pathway in cancer, basal cell carcinoma, apoptosis, induction of programmed cell death, regulation of transcription (DNA-templated), intracellular signal transduction, and regulation of peptidase activity. In particular, we found that the levels of H3K4Me3 at the promoters of SUSD2, CCND2, BCL2AI, and TMEMIS8 are significantly altered in A549, NCI-H1975, NCI-H1650, and NCI-H2228 cells, when treated with JFK. Collectively, these findings provide the first evidence that the anticancer activity of JFK involves modulation of histone modification at many cancer-related gene loci.

#### 1. Introduction

Chromatin is the macromolecular complex of DNA and histone proteins that provides the scaffold for packaging the eukaryotic genome [1, 2]. Histones H2A, H2B, H3, and H4 are the basic components of nucleosomes, which form the fundamental unit of chromatin [3, 4]. Chemical modifications to the histones alter chromatin structure and regulate gene expression by altering noncovalent interactions within and between nucleosomes [2, 5]. H3K4Me3 is an active histone modification which is positively associated with gene expression [3, 6]. Previous studies have shown that the levels of H3K4Me3 modification are closely associated with the development, treatment, and diagnosis of

disease [7-9]. Chromatin immunoprecipitation followed by sequencing (ChIP-seq) has been developed to systematically characterize the contribution of epigenetic regulation in various biological processes via genome-wide profiling of various chemical modifications of histone proteins and genomic DNA methylation [10].

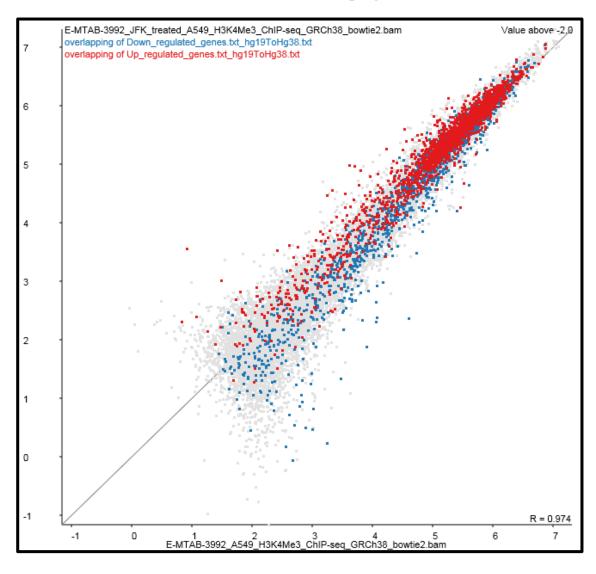
Lung cancer has become the leading cause of cancerrelated deaths worldwide [11]. Overall, only 16.8% of patients with lung cancer survive five years after their first definite diagnosis, mainly as a consequence of uncontrollable cell proliferation or tumor metastasis [12, 13]. Although various therapeutic interventions, including surgery, chemotherapy, and radiotherapy, have been developed to prolong the survival time of patients, drug side effects, pain, and emaciation

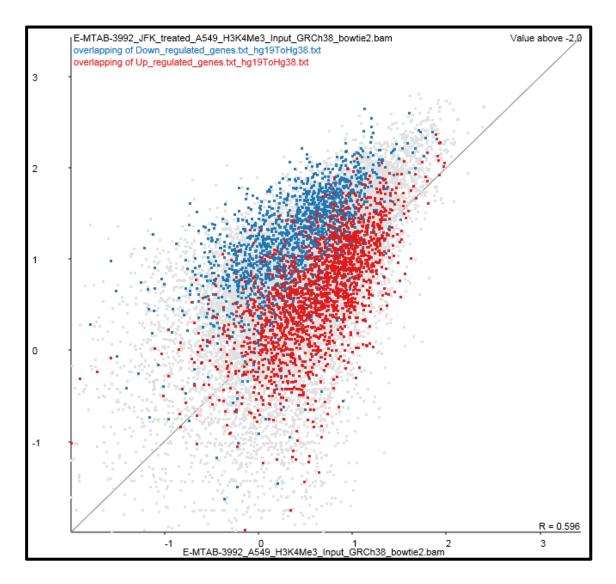
<sup>&</sup>lt;sup>1</sup>Shanghai Center for Systems Biomedicine, School of Biomedical Engineering, State Key Laboratory on Oncogene and Bio-ID Center, Shanghai Jiao Tong University, 800 Dongchuan Road, Shanghai 200240, China

<sup>&</sup>lt;sup>2</sup>Tumor Institute of Traditional Chinese Medicine, Longhua Hospital, Shanghai University of Traditional Chinese Medicine, 725 South Wanping Road, Shanghai 200032, China

<sup>3</sup> College of Life Science, Northwest University, 229 Taibai Road, Xi'an 710069, China

# Bias or Biology?





ChIP

Input

# What can you do?

 Think about whether you're likely to have expected biases in your experiment.

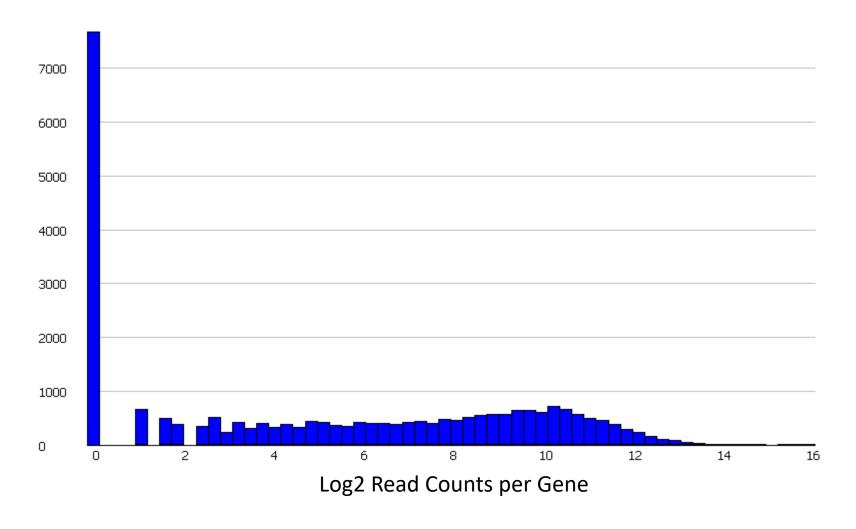
- Look for unexpected biases.
  - Sometimes the bias is the interesting biology

 Use custom backgrounds during Gene Set Analysis to help minimise bias (if a tool supports it)

## Using a background list can make a huge difference

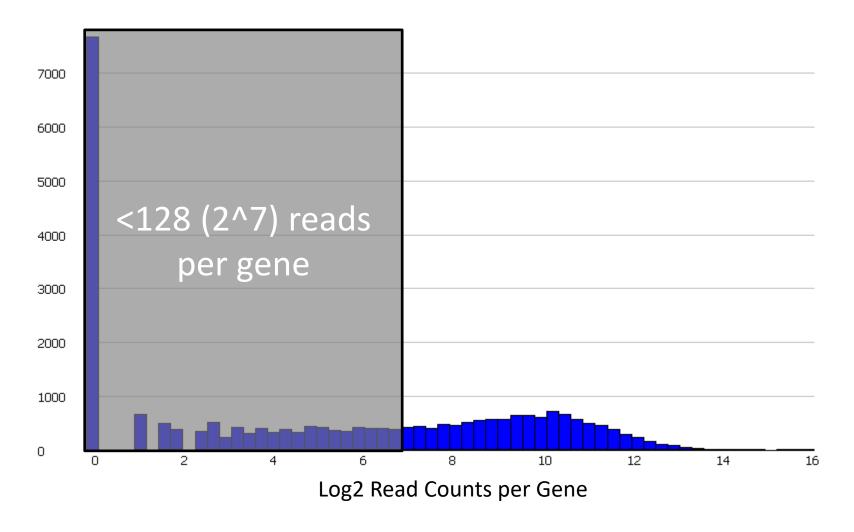
- What genes were you likely to see?
  - Some are technically impossible
    - Membrane proteins in LC-MS
    - Small-RNA in RNA-Seq
  - Some are much less likely
    - Unexpressed or low expressed in RNA-Seq
    - Unmappable in ChIP-Seq
    - Low CpG content in BS-Seq
- Make a list of what you could have seen, and set that as the background.

# **Expressed Genes**



26,127 Genes Measured

# **Expressed Genes**

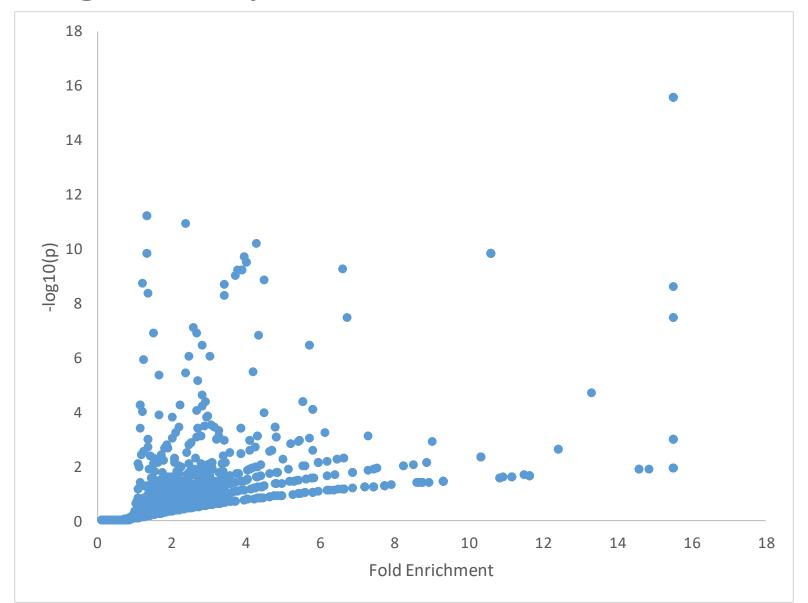


10,378 Genes Realistically Measured (40%)

# Statistical biases affect gene sets too

- Fisher's test is powered by
  - Magnitude of change
  - Observation level
- Big lists have more power to detect change
- Small lists are very difficult to detect
- Some tools allow you to exclude the largest gene set categories. We often use categories with between 50 500 genes in to get power and specificity
- Always look at the enrichment and the p-value when deciding what is interesting

# Fold Change and p-value



## Other biases: Random Genomic Positions

- Relating genomic positions to genes
- Find closest gene
  - Synapse, Cell Junction, postsynaptic membrane (p=8.9e-12)
  - Membrane (p=4.3e-13)
- Find overlapping genes
  - Plekstrin homology domain (p=1.8e-7)
  - Ion transport (p=7.1e-7)

Creating a background list with the same biases as your hit list will alleviate the artefacts.

# Stuff which turns up more than it should...

- Did a trawl through GEO RNA-Seq datasets
  - Downloaded pairs of samples which are supposed to be biological replicates
  - Found changing genes
  - Ran GO searches
- Many gene sets give hits. Some categories turn up very often
  - Ribosomal
  - Cytoskeleton
  - Extracellular
  - Secreted
  - Translation



www.bioinformatics.babraham.ac.uk/projects/goliath/

#### Welcome to GOliath

Select species	Homo_Sapiens/Dec_18
Min Category Size	50
Max Category Size	500
Gene List	Background List (optional)
Paste Gene Names here  .:i	Paste Gene Names here
Use example genes	

Analyse my list



www.bioinformatics.babraham.ac.uk/projects/goliath/

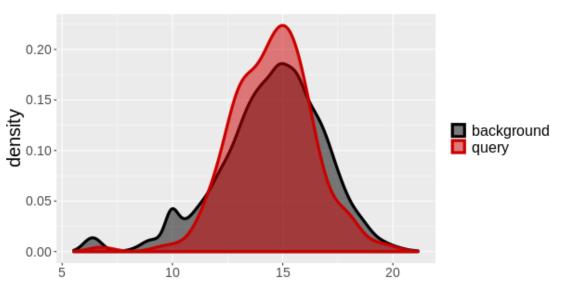
Results for job Course test

Results Table			Prop	erties		Biase	s
Copy CSV Excel Prin	nt		Hit table				
Gene Set	Source	Query count	Background count	Category size	FDR	Enrichment	Potential bias
HALLMARK TNFA SIGNALING VIA NFKB	MSIGDB C2 HALLMARK TNFA SIGNALING VIA NFKB	17	200	200	1.384e-07	9.174	public_data
SIGNALING BY INTERLEUKINS	REACTOME R-HSA- 449147.11	20	461	461	4.195e-05	4.683	high_transcripts
POSITIVE REGULATION OF CYTOKINE PRODUCTION	GOBP GO:0001819	16	355	355	0.0005164	4.865	public_data
HALLMARK IL2 STAT5 SIGNALING	MSIGDB C2 HALLMARK IL2 STAT5 SIGNALING	12	200	200	0.0009546	6.476	public_data
HALLMARK APOPTOSIS	MSIGDB C2 HALLMARK APOPTOSIS	10	160	160	0.003579	6.746	
APOPTOSIS	WIKIPATHWAYS 20190910 WP254 HOMO SAPIENS	8	87	87	0.003579	9.925	
REGULATION OF CYTOKINE SECRETION	GOBP GO:0050707	10	154	154	0.003579	7.009	
REGULATION OF INTERLEUKIN-6 PRODUCTION	GOBP GO:0032675	8	101	101	0.007389	8.549	
HALLMARK INFLAMMATORY RESPONSE	MSIGDB C2 HALLMARK INFLAMMATORY RESPONSE	10	200	200	0.008619	5.397	public_data
HALLMARK ALLOGRAFT REJECTION	MSIGDB C2 HALLMARK ALLOGRAFT REJECTION	10	200	200	0.008619	5.397	public_data
Search gene set	Search source	min query	min bg	min size	min FDR	min enrichment	Search bias
		max query	max bg	max size	max FDR	max enrichment	

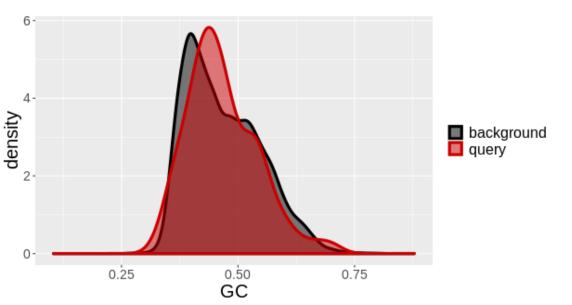
# Checking for unexpected biases

- Do my hits look different from non-hits in factors which should be unrelated
  - Sequence composition
  - Genomic position
  - Gene Length
  - Number of splice variants
  - etc
- If a bias exists then is this the actual link between genes? If not then can I fix this by improving my background list?

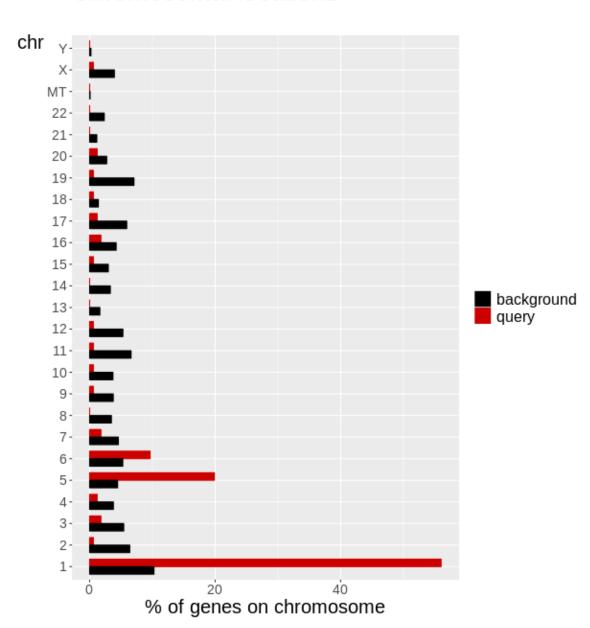
### Gene lengths



### GC content of genes

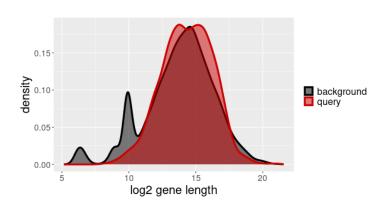


### **Chromosomal locations**

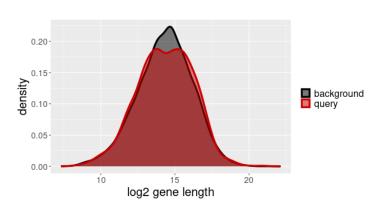


# Custom backgrounds can make a difference

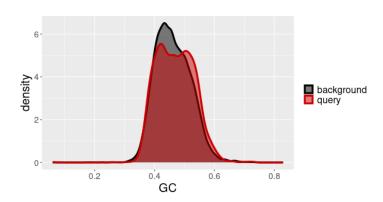
#### Gene lengths



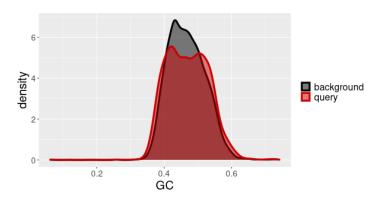
#### Gene lengths



#### GC content of genes



#### GC content of genes



# Custom backgrounds can make a difference

## Top hits without correction

**PLURINETWORK** 

POSITIVE REGULATION OF VASCULATURE DEVELOPMENT POSITIVE REGULATION OF ANGIOGENESIS

HALLMARK E2F TARGETS

CHROMOSOME, CENTROMERIC REGION

DNA REPAIR

NEGATIVE REGULATION OF CELLULAR AMIDE METABOLISM

POSITIVE REGULATION OF ENDOTHELIAL CELL MIGRATION

NUCLEAR CHROMOSOME SEGREGATION

PID INTEGRIN1 PATHWAY

## Top hits with correction

POSITIVE REGULATION OF VASCULATURE DEVELOPMENT

POSITIVE REGULATION OF ANGIOGENESIS

PID INTEGRIN1 PATHWAY

BETA1 INTEGRIN CELL SURFACE INTERACTIONS

INTEGRIN BINDING

ASSEMBLY OF COLLAGEN FIBRILS

NABA ECM REGULATORS

POSITIVE REGULATION OF ENDOTHELIAL CELL MIGRATION

RECEPTOR LIGAND ACTIVITY

STRIATED MUSCLE TISSUE DEVELOPMENT

# **Avoiding Biases**

- Create a custom background if applicable
  - Should contain all genes which \*could\* have been in your hit list
  - May be a compromise, but it's better than nothing
  - Will limit which tools you can run

- Filter your tested gene sets
  - Remove large over powered sets, or sets which are too small to achieve significance (~50 to ~500 is generally about right)
  - Check the hit gene sets for matches to known problematic sets

# Programmatic Gene List Analysis in R

Simon Andrews

simon.andrews@babraham.ac.uk



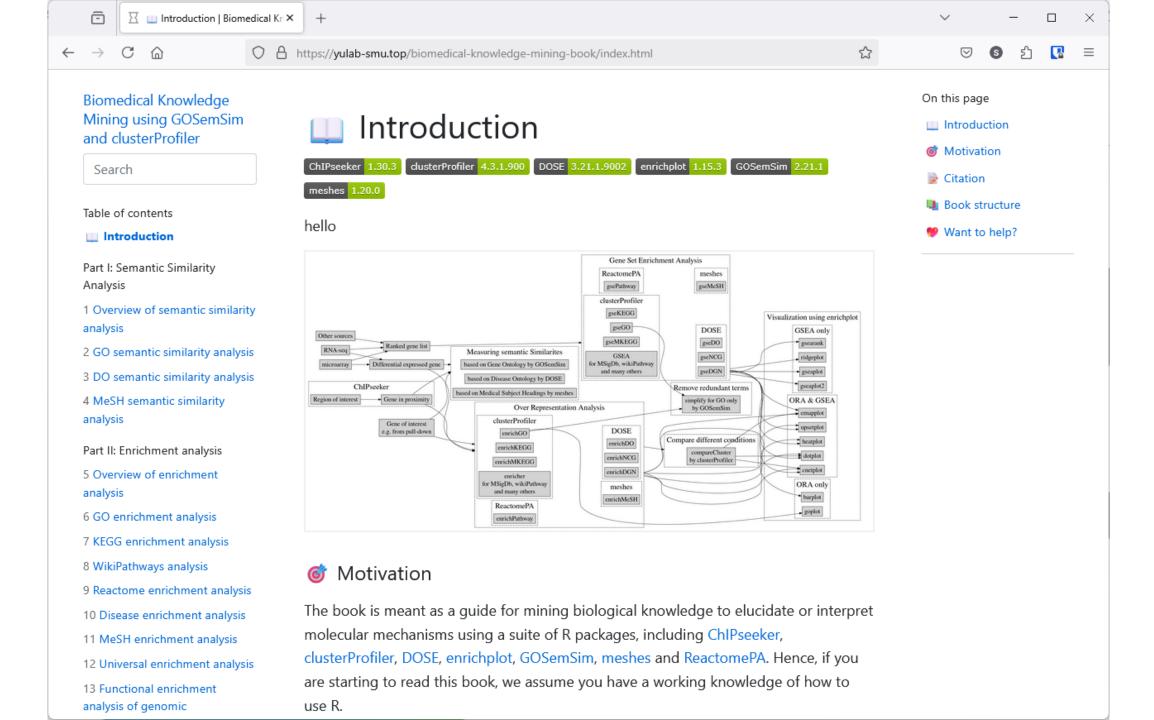


## clusterProfiler

This is the **released** version of clusterProfiler; for the devel version, see <u>clusterProfiler</u>.

## A universal enrichment tool for interpreting omics data

- Categorical or quantitative enrichment
- Multi-species support



## **Genome Annotations**

## org.Hs.eg.db

This is the released version of org.Hs.eg.db; for the devel version, see org.Hs.eg.db.

## Genome wide annotation for Human

## GO.db

This is the **released** version of GO.db; for the devel version, see GO.db.

## A set of annotation maps describing the entire Gene Ontology

org.Ag.eg.db	Anopheles
org.At.tair.db	Arabidopsis
org.Bt.eg.db	Bovine
org.Cf.eg.db	Canine
org.Gg.eg.db	Chicken
org.Pt.eg.db	Chimp
org.EcK12.eg.db	E coli K12
org.Dm.eg.db	Fly
org.Hs.eg.db	Human
org.Pf.plasmo.db	Malaria
org.Mm.eg.db	Mouse
org.Ss.eg.db	Pig
org.Rn.eg.db	Rat
org.Mmu.eg.db	Rhesus
org.Ce.eg.db	Worm
org.Xl.eg.db	Xenopus
org.Sc.sgd.db	Yeast
org.Dr.eg.db	Zebrafish

# Installation and loading

```
install.packages("BiocManager")
BiocManager::install("clusterProfiler")
BiocManager::install("org.Hs.eg.db")
library(clusterProfiler)
library(org.Hs.eg.db)
library(tidyverse)
```

# Different types of enrichment analysis

- Categorical enrichment
  - enrichGO
  - enrichPathway

- Quantitative enrichment
  - gseGO
  - gsePathway

# Categorical Enrichment Analysis



# Categorical Gene Ontology Enrichment

- Input requirements
  - Vector of hit gene ids
  - Keytype for the ids
  - Vector of all analysed gene ids

#### > keytypes(org.Hs.eg.db)

```
"ACCNUM"
                     "ALIAS"
                                                     "ENSEMBLPROT"
                                     "ENSEMBL"
                                                                      "ENSEMBLTRANS"
     "ENTREZID"
                     "ENZYME"
                                     "EVIDENCE"
                                                     "EVIDENCEALL"
                                                                      "GENENAME"
    "GENETYPE"
                     "GO"
                                     "GOALL"
                                                     "IPI"
                                                                      "MAP"
     "OMIM"
                     "ONTOLOGY"
                                     "ONTOLOGYALL"
                                                     "PATH"
                                                                      "PFAM"
[16]
                     "PROSITE"
                                     "REFSEO"
                                                     "SYMBOL"
                                                                      "UCSCKG"
    "PMID"
    "UNIPROT"
```

# Picking the correct keytype

```
select(
  org.Hs.eg.db,
  keys=keys(org.Hs.eg.db)[1:10],
  columns=c("ENSEMBL","SYMBOL","GENENAME")
)
```

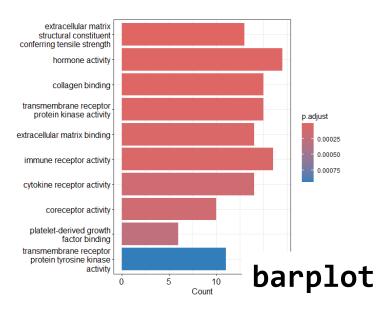
<b>ENTREZID</b>	ENSEMBL	SYMBOL	GENENAME
1	ENSG00000121410	A1BG	alpha-1-B glycoprotein
2	ENSG00000175899	A2M	alpha-2-macroglobulin
3	ENSG00000291190	A2MP1	alpha-2-macroglobulin pseudogene 1
9	ENSG00000171428	NAT1	N-acetyltransferase 1
10	ENSG00000156006	NAT2	N-acetyltransferase 2
12	ENSG00000196136	SERPINA3	serpin family A member 3
13	ENSG00000114771	AADAC	arylacetamide deacetylase
14	ENSG00000127837	AAMP	angio associated migratory cell protein
15	ENSG00000129673	AANAT	aralkylamine N-acetyltransferase

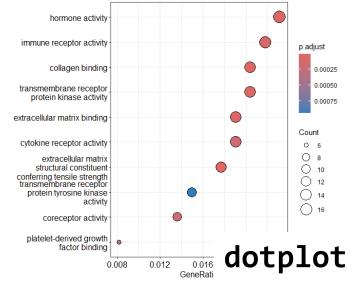
# Running a categorical enrichment analysis

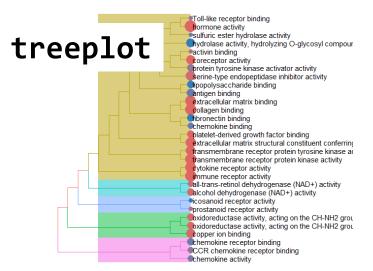
```
enrichGO(
   up_genes,
   OrgDb = org.Mm.eg.db,
   keyType = "SYMBOL",
   universe = background_genes,
   ont = "BP",
   minGSSize = 10,
   maxGSSize = 100,
   readable = TRUE
) -> enrichgo_results
```

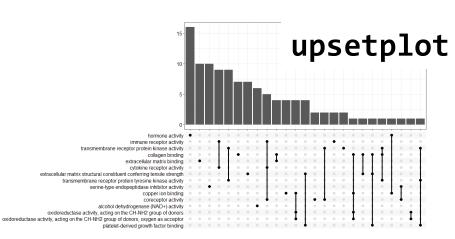
```
enrichgo_results@result |>
  as_tibble()
```

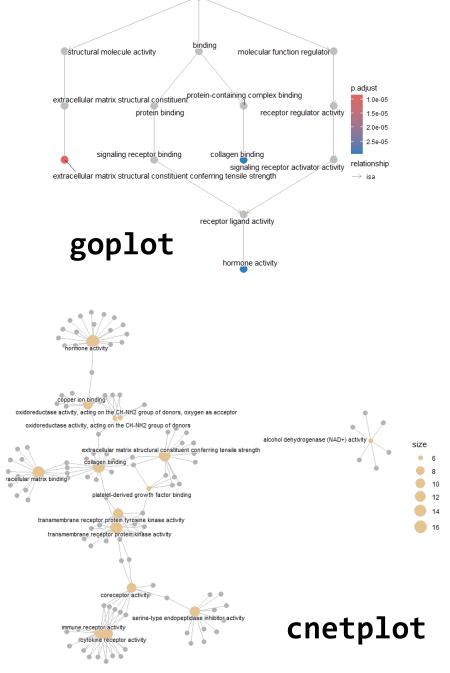
ID <chr></chr>	Description <chr></chr>	GeneRatio <chr></chr>	BgRatio <chr></chr>	FoldEnrichment <dbl></dbl>	zScore <dbl></dbl>	pvalue <dbl></dbl>	<b>p.adjust</b> <dbl></dbl>	<b>qvalue</b> <dbl></dbl>
GO:0030199	collagen fibril organization	22/734	69/17422	7.567903	11.464270	3.666706e-14	1.249980e-10	1.017800e-10
GO:1903053	regulation of extracellular matrix organization	18/734	65/17422	6.572962	9.440344	1.102615e-10	1.879407e-07	1.530313e-07
GO:0072676	lymphocyte migration	19/734	81/17422	5.567632	8.641298	7.579679e-10	8.613041e-07	7.013197e-07
GO:0072678	T cell migration	15/734	56/17422	6.357775	8.421919	6.582620e-09	5.610038e-06	4.567992e-06
GO:1903487	regulation of lactation	7/734	10/17422	16.614986	10.358568	2.460222e-08	1.677380e-05	1.365812e-05
GO:2000401	regulation of lymphocyte migration	14/734	57/17422	5.829820	7.659729	7.008763e-08	3.549840e-05	2.890469e-05
GO:0031342	negative regulation of cell killing	10/734	27/17422	8.790998	8.496600	7.289200e-08	3.549840e-05	2.890469e-05
GO:2000404	regulation of T cell migration	12/734	42/17422	6.781627	7.867419	9.866697e-08	3.969711e-05	3.232350e-05
GO:0030168	platelet activation	18/734	97/17422	4.404562	7.051685	1.048032e-07	3.969711e-05	3.232350e-05
GO:0050830	defense response to Gram-positive bacterium	16/734	81/17422	4.688532	6.978169	2.173487e-07	7.409417e-05	6.033142e-05



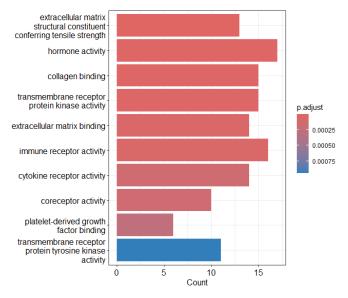


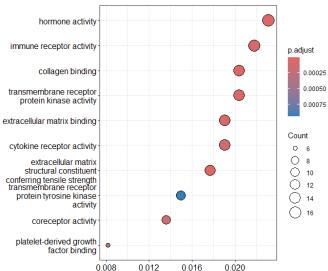






molecular\_function





GeneRatio

```
barplot(enrichgo_results, showCategory = 10)
dotplot(enrichgo_results, showCategory = 10)
```

```
enrichgo_results |>
    dotplot(
        x="FoldEnrichment",
        showCategory=20
)
```

```
enrichgo_results |>
    dotplot(
        x="Count",
        showCategory=20
)
```

#### **GeneRatio**

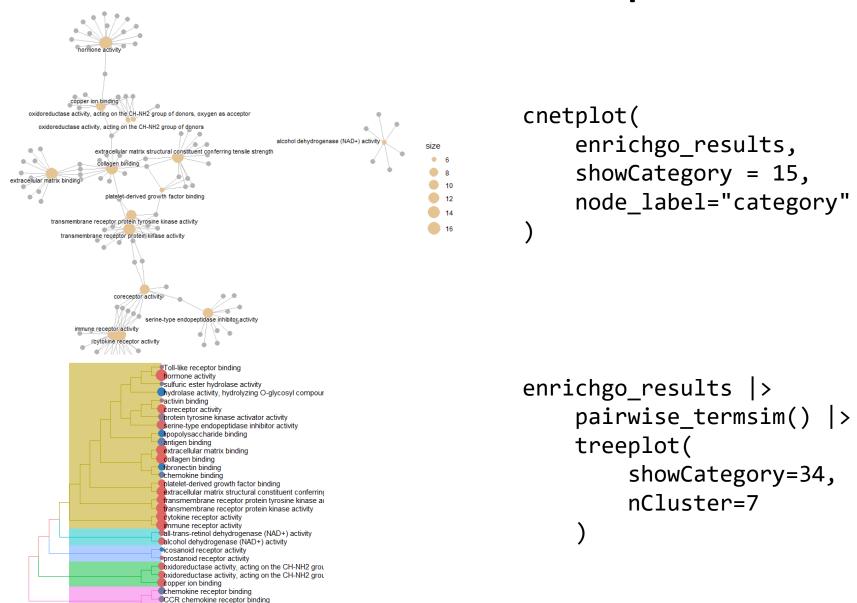
Fraction of background in hit list

#### Count

Absolute number in hit list

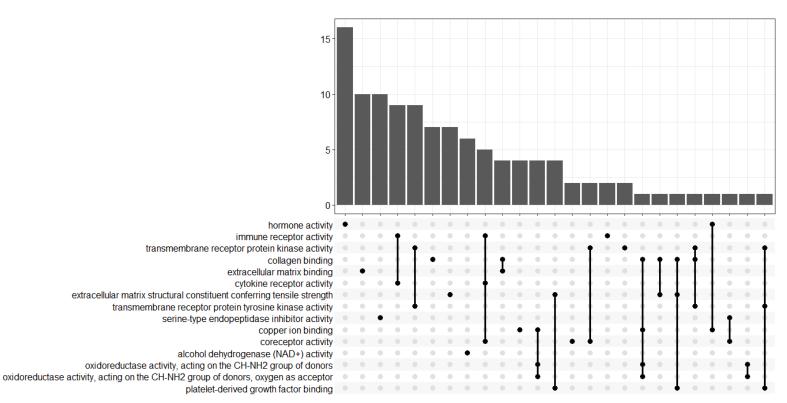
#### **Fold Enrichment**

Fraction in hits over fraction in background



chemokine activity

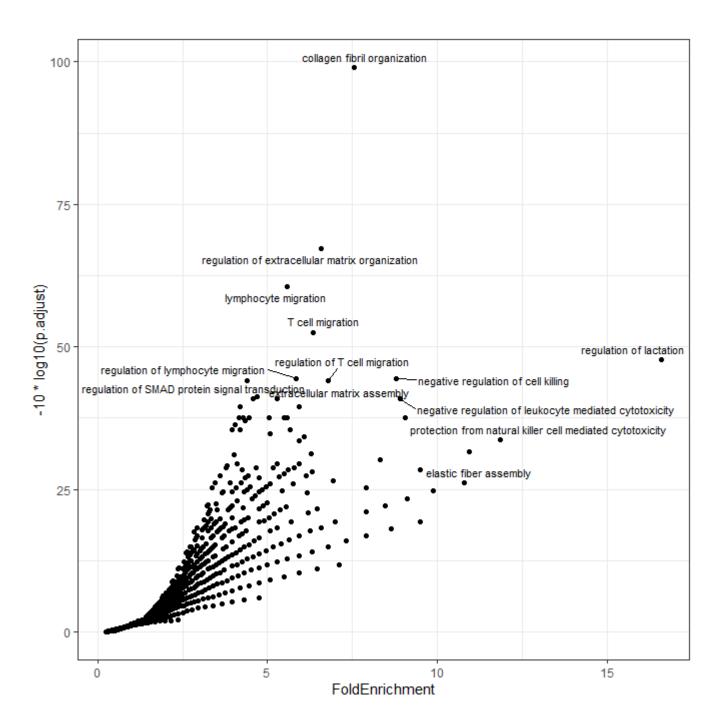
- all
- none
- category
- gene



```
upsetplot(
     enrichgo_results,
     n=15
)
Note: *not* showCategory in this plot
```

## **Custom Visualisation**

```
enrichgo_results@result |>
  as_tibble(rownames="GOID") |>
  ggplot(aes(
    x=FoldEnrichment,
   y=-10*log10(p.adjust),
    label=Description
  geom point() +
  ggrepel::geom_text_repel(
    data = . %>% filter(
      FoldEnrichment > 5 &
      (-10*log10(p.adjust))>20
    size=3
```



# Categorical Enrichment with R Exercise



# Quantitative Enrichment Analysis



## Named vectors

```
c(1,5,3,6) \rightarrow my data
my data
[1] 1 5 3 6
c("simon", "sarah", "jo", "hayley") -> names(my data)
my data
 simon sarah jo hayley
```

GSEA input is a gene named vector of sorted (high to low) values

# Running a quantitative enrichment analysis

Gene <sup>‡</sup>	Chromosome <sup>‡</sup>	Start <sup>‡</sup>	End <sup>‡</sup>	ID <sup>‡</sup>	Description	Type <sup>‡</sup>	P-value	FDR <sup>‡</sup>	<b>ZScore</b>
Xkr4	1	3276124	3741721	ENSMUSG00000051951	X-linked Kx blood group related 4 [Source:MGI Symbol;Acc:	gene	3.604699e-02	0.455440968	1.798524261
Rp1	1	4069780	4479464	ENSMUSG00000025900	retinitis pigmentosa 1 (human) [Source:MGI Symbol;Acc:MG	gene	3.510687e-01	0.645575404	0.382436782
Sox17	1	4561154	4567577	ENSMUSG00000025902	SRY (sex determining region Y)-box 17 [Source:MGI Symbol;	gene	8.803802e-02	0.587544382	-1.352936149
Mrpl15	1	4843429	4855962	ENSMUSG00000033845	mitochondrial ribosomal protein L15 [Source:MGI Symbol;A	gene	3.934556e-01	0.645575404	0.270323783
Lypla1	1	4878011	4918633	ENSMUSG00000025903	lysophospholipase 1 [Source:MGI Symbol;Acc:MGI:1344588]	gene	3.866758e-01	0.645575404	0.287993759
Tcea1	1	4928037	4968132	ENSMUSG00000033813	transcription elongation factor A (SII) 1 [Source:MGI Symbol;	gene	3.479390e-01	0.645575404	0.390890777
Rgs20	1	4979799	5140508	ENSMUSG00000002459	regulator of G-protein signaling 20 [Source:MGI Symbol;Acc	gene	4.381771e-02	0.480191737	1.708004951
Atp6v1h	1	5140241	5232752	ENSMUSG00000033793	ATPase, H+ transporting, lysosomal V1 subunit H [Source:M	gene	3.373898e-01	0.645575404	0.419597268
Oprk1	1	5658689	5676354	ENSMUSG00000025905	opioid receptor, kappa 1 [Source:MGI Symbol;Acc:MGI:97439]	gene	1.760258e-01	0.643769562	0.930617332
Npbwr1	1	5983926	5987617	ENSMUSG00000033774	neuropeptides B/W receptor 1 [Source:MGI Symbol;Acc:MGI	gene	3.946832e-01	0.645575404	0.267133325

```
kat6ab_zscore |>
   arrange(desc(ZScore)) |>
   distinct(Gene, .keep_all = TRUE) -> data

data |>
   pull(ZScore) -> zscore_data

data$Gene -> names(zscore_data)
```

Fabp7 7.232035 Gm3764 6.434833 Reln 6.374436 Gfap 6.281050

Gm35040 C130071C03Rik 5.209227 5.187987

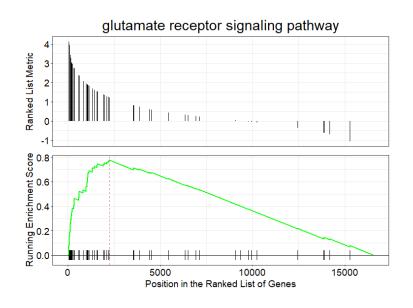
# Running a quantitative enrichment analysis

```
gseGO(
  zscore_data,
  ont="BP",
  OrgDb = "org.Mm.eg.db",
  keyType = "SYMBOL",
  minGSSize = 10,
  maxGSSize = 100
) -> gsego_result
```

```
gsego_result@result |>
  as_tibble()
```

ID <chr></chr>	Description <chr></chr>	setSize <int></int>	enrichmentScore <dbl></dbl>	NES <dbl></dbl>	<b>pvalue</b> <dbl></dbl>	p.adjust <dbl></dbl>
GO:0030199	collagen fibril organization	69	-0.6965226	-2.952459	1.000000e-10	4.995000e-08
GO:0007215	glutamate receptor signaling pathway	57	0.7761472	2.374072	1.000000e-10	4.995000e-08
GO:0099172	presynapse organization	66	0.7500618	2.348110	1.000000e-10	4.995000e-08
GO:0099505	regulation of presynaptic membrane potential	33	0.8316328	2.327325	1.000000e-10	4.995000e-08
GO:0099054	presynapse assembly	53	0.7530821	2.291861	1.000000e-10	4.995000e-08
GO:0051966	regulation of synaptic transmission, glutamatergic	83	0.7158265	2.287267	1.000000e-10	4.995000e-08
GO:0099174	regulation of presynapse organization	43	0.7731902	2.272698	1.000000e-10	4.995000e-08
GO:0019226	transmission of nerve impulse	100	0.6740946	2.209081	1.000000e-10	4.995000e-08
GO:1903539	protein localization to postsynaptic membrane	76	0.6860463	2.185224	1.000000e-10	4.995000e-08
GO:0062237	protein localization to postsynapse	79	0.6799646	2.170268	1.000000e-10	4.995000e-08

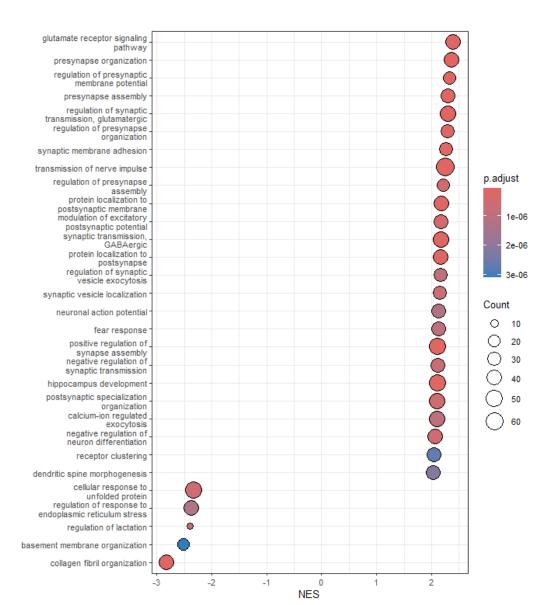
## **GSEA Visualisation**



```
gseaplot(
  gsego_result,
  geneSetID = 2,
  title = gsego_result$Description[2]
)
```

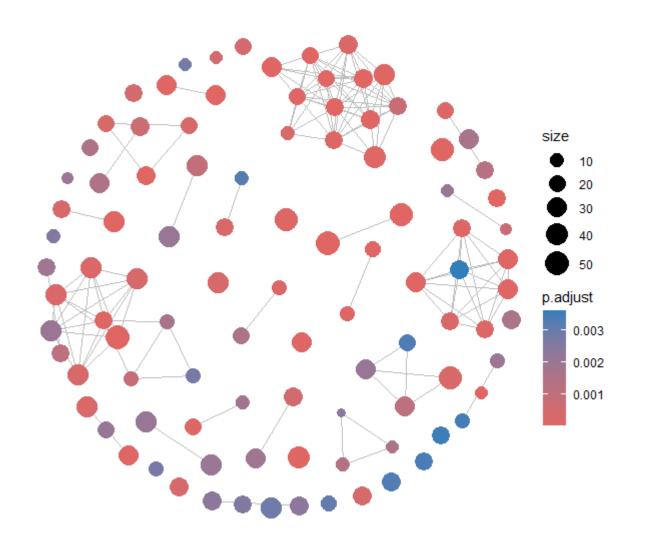
```
gseaplot2(
  gsego_result,
  geneSetID = 2,
  title = gsego_result$Description[2]
)
```

## **GSEA Visualisation**



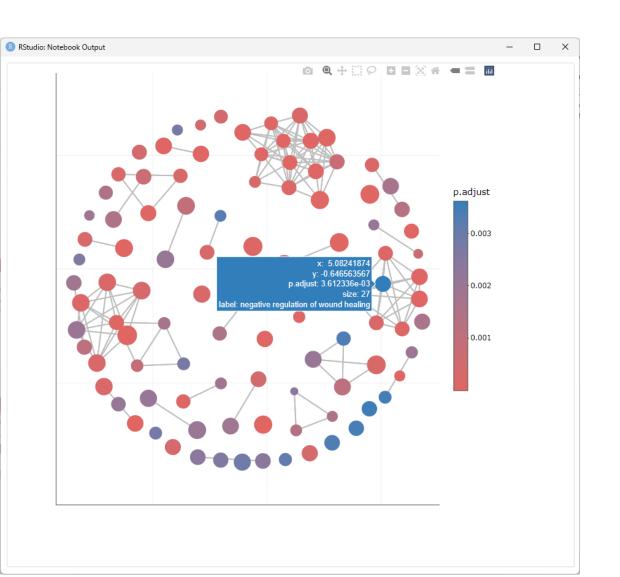
```
dotplot(
   gsego_result,
   x="NES",
   showCategory=30
)
```

## **GSEA Visualisation**



```
gsego_result |>
  filter(NES < 0) |>
  pairwise_termsim() |>
  emapplot(
    min_edge=0.5,
    showCategory=100,
    node_label="none"
)
```

# Interactive visualisation (kludge!)



```
gsego_result |>
  filter(NES < 0) |>
  pairwise_termsim() |>
  emapplot(min_edge=0.5,
      showCategory=100,
      node_label="category") -> temp

temp$layers[[2]]$mapping$label =
  temp$data$label

plotly::ggplotly(temp, width = 800, height=800)
```

# Quantitative Enrichment with R Exercise

